

Protective Effect of Ethanolic Extract of *Zingiber Officinale* Against Mercuric Chloride Induced Renal Toxicity In Rats

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Abstract

The current study was designed to investigate the possible protective effects of ethanolic extract of *Zingiber officinale* on renal damage induced by mercuric chloride intoxication in rats. Forty albino adult rats (180±10g, n=10 per group) were administered HgCl₂ (3mg/kg, Ip, 3 times weekly). The treatment of *Z. officinale* extract (200 mg orally for 8 weeks. HgCl₂ administration altered the body weight, kidney weight, chemical parameters of creatinine, urea, and MDA. Body, kidney weight the biochemical and histopathological findings of this study significantly (P≤0.05) showed a protective effect of a *Z. officinale* extract against HgCl₂ intoxication. In conclusion, *Z. officinale* extracts may be an ideal choice against kidney damage induced by HgCl₂.

Keywords: mercuric chloride, *Zingiber* extract, histopathology, kidney.

Introduction

Mercuric chloride accumulates mostly in rat liver and kidney as these organs involved in the detoxification and excretion of foreign materials¹. Mercury contamination of the environment continues to be a concern and a major source of this contamination is from human activities by industrial waste which releases mercury into the environment². The renal cortex and liver are considered to be the most susceptible organs affected which leads to functional impairment (3). Several studies were focused on the role of free radicals and oxidants due to mercuric toxicity affect the proximal convoluted tubules injury, which suggests that the nephron plays an important role in the active transport of this heavy metal^{3,4,5,6,7,8,9}

Natural herbaceous plant *Zingiber officinale* Roscoe is one of the most common food-flavoring spices used worldwide⁷. For many years, several pharmacological properties of ginger, such as anti-inflammatory, analgesic, gastrointestinal regulating agent, antioxidant and antimicrobial properties have been identified^{8,9,10}. Efforts are now being directed in obtaining drugs with different chemical features since many modern drugs originated from plants the investigation of the chemical composition of traditional medicinal plants could lead to the development of new drugs. The study therefore, investigated the possibility of utilizing the ethanolic extract of *Z. officinale* in the treatment of HgCl₂ induced nephrotoxicity.

Method

Experimental Design: Forty healthy adult albino rats 180±19g, were separated into four groups of ten rats for each group. eight weeks the animals kept in suitable cages and was feeder standard diet and water ad libitum. The control group received distilled water (10ml/kg/ intraperitoneal (Ip) the second group-administered three times per week (ip) injection of HgCl₂ at a dose of 3mg/

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kg. Third group, treated with [HgCl₂(3mg/kg) plus (200mg/daily orally)] of *Z officinale* extract, and the fourth group treated with *Z officinale* extract (200mg/daily orally).

Sample Preparation: At the end of 8 weeks, all animals were scarified the body and kidney weight is measured. Clear sera were collected and stored at -20°C for biochemical assay. The kidneys was rapidly excised and stored in 10% formalin solution and processed for histological examination.

Determination of biochemical and histomorphometric parameters: Serum creatinine, urea and lipid peroxide contents were measured in all rats involved in this study using standard laboratory method.

Body and kidney weights were estimated at the end of 8 weeks. Under light microscopy the number and the diameter of (pct) were measured beside the histological examination.

Statistical Analysis: The data were expressed as means \pm standard deviation ($X \pm SD$). ANOVA (analysis of variance) to compare between more than two groups of numerical (parametric) data followed by post hoc Tukey test. A P value <0.05 was considered statistically significant.

Results

Effect of Zingiber officinale extract treatment on body and kidney weights: Table 1 showed a significant decrease in (b.w) and a significant increase in (k.w) of the rats among the HgCl₂ group ($P \leq 0.05$). While the (k.w) of the rats treated with extract of *Z officinale* showed a significant decrease when compared with rats among the HgCl₂ group ($P \leq 0.05$). No significant difference in the body and kidney weight between (*Z officinale* + HgCl₂) group and the control group.

Effect of Zingiber officinale extract treatment on the diameter and number of (pct): Means diameter of

(pct) shows a significant increase in HgCl₂ treated group compared with other groups ($P \leq 0.05$), while the number of (pct) showed a significant decrease in HgCl₂ compared with other treated groups ($P \leq 0.05$). Table 3 showed a significant decrease in the diameter of (pct) among rats treated with *Z officinale* + HgCl₂ group ($P \leq 0.05$), also showed a significant increase in the number of the (pct) of *Z officinale* + Hgcl₂ group ($P \leq 0.05$). No significant difference in the diameter of (pct), and the number of (pct) of the *Z officinale* extract and control group.

Table 1: Effect of Zingiber officinale extract treatment on body and kidney weights

Groups	Bodyweight (gm)	Kidney Weight (gm)
Control	190.50 \pm 6.123 (A)	1.397 \pm 0.013 (C)
HgCl ₂	157.83 \pm 2.041 (C)	4.055 \pm 0.181 (A)
HgCl ₂ +Z officinale	171.50 \pm 5.205 (B)	2.220 \pm 0.216 (B)
Z officinale	189.67 \pm 5.887 (A)	1.406 \pm 0.019 (C)

Values are expressed as mean \pm SD of 10 rats in each group. Significantly different from Group HgCl₂ ($P \leq 0.05$). Capital letters denote differences between the groups.

Table 3: Effect of Zingiber officinale extract treatment on the diameter and number of (pct)

Groups	The Diameter of (pct)	No. of (pct)
Control	41.373 \pm 0.329 (C)	17.728 \pm 0.746 (A)
HgCl ₂	53.353 \pm 0.654 (A)	14.725 \pm 0.502 (C)
Z officinale + HgCl ₂	44.893 \pm 1.0731 (B)	16.415 \pm 0.521 (B)
Z officinale	41.165 \pm 0.245 (C)	18.036 \pm 0.934 (A)

Values are expressed as mean \pm SD of 10 rats in each group. Significantly different in Group Hgcl₂, ($P \leq 0.05$). Capital letters denote differences between the groups, $P \leq 0.05$ vs. control.

Effect of Zingiber officinale extract treatment on creatinine, urea and MDA levels: Creatinine, urea and MDA levels shows significant elevate in the HgCl₂ group of rats compared with the control group ($p < 0.05$). While the *Z officinale* + HgCl₂ group rats shows a significant reduction in the serum creatinine and urea levels ($P \leq 0.05$) (table 2).

Table 2: Effect of Zingiber officinale extract treatment on creatinine, urea and MDA levels

Groups	Creatinine	Urea	MDA
Control	127.96 \pm 0.076 (C)	58.045 \pm 1.413 (C)	4.365 \pm 0.199 (C)
HgCl ₂	437.77 \pm 24.013 (A)	149.08 \pm 1.652 (A)	7.908 \pm 0.263 (A)
HgCl ₂ +Z officinale	244.67 \pm 14.684 (B)	87.848 \pm 1.889 (B)	5.063 \pm 0.398 (B)
Z officinale	123.77 \pm 0.537 (C)	55.400 \pm 0.530 (C)	4.410 \pm 0.197 (C)

Values are expressed as mean \pm SD of 10 rats in each group. Significantly different from Group HgCl₂ ($P \leq 0.05$). Capital letters denote differences between groups.

Morphological Examination: The kidneys of rats of the *Z officinale* extract, *Z officinale* + HgCl_2 groups showed bean-shaped, reddish-brown in color with smooth and possess convex and concave borders nearly normal (fig. 1a,b & d), while the kidney of rats of the HgCl_2 group was pale, swollen and enlarged (fig. 1c).

Histological Examination: Figure 2a: show histopathological findings from H & E stained kidney sections. In the control group rats, no glomerular or pathological abnormalities. However, HgCl_2 group rats showed multiple pathological changes, which include dilated of Bowman's capsules, atrophy glomerulus tuft, vacuolated of epithelial cells and cast formation with the presence of interstitial exudates. Also, a glomerular cell cast of renal tubules proximal dense nuclear of epithelial cell, hyper atrophy of other cells, a hemorrhagic area with a mild aggregation of some inflammatory cells beside a swelling of epithelial cells as shown in (fig. 2b). Rat kidneys treated with *Z officinale* + HgCl_2 group showed normal renal tubules and nearly normal epithelial cells, with very mild degeneration, while normal kidney tissues appear in (fig. 2c) rats.



Fig. 1: a, c & d. (normal, *Z officinale* & HgCl_2 + *Z officinale* extract groups) showed normal gross appearance kidney. b, HgCl_2 group showing pale and swollen kidney.

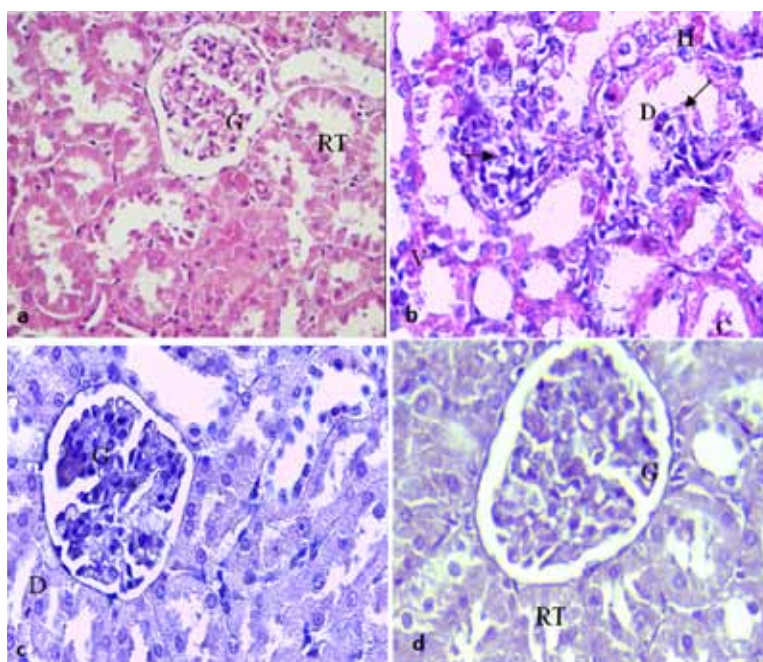


Fig. 2: a. Control group, showing normal glomeruli (G) and normal renal tubules (RT). b, HgCl_2 group showing dilated of Bowman's capsules (D), glomeruli tuft atrophy (\rightarrow) vacuolated of epithelial cells (V) and glomerulus cells cast of renal tubules (C), hemorrhagic area (H). c, *Zingiber officinale* + HgCl_2 group showed nearly normal renal tubules with normal epithelial cells, and very mild degeneration (D). d, *Zingiber officinale* group showed normal glomerulus tuft, normal renal tubules and normal glomeruli of Bowman capsule H & E400X.

Discussion

The pathogenesis of nephrotoxicity of mercury is due to binding of mercuric ions with sulfur, like a thiol group of amino acids, which transfer mercury ion via sodium ion channels to the kidney tubules, causing in accumulation in the renal proximal tubular cells and increasing free radicals formation and damaging renal tubules^{4,11,12}. So the current study was undertaken to determine whether Zingiber can reduce or prevent HgCl₂ induced renal damage by examining different biochemical and histological parameters related to kidney function of intoxicated and treated rats.

Reduce of the body weight may be resulting from being an interruption of HgCl₂ in the absorption and metabolism of feed nutrients essential for health¹³. Mercury is capable of damaging the organism in many ways because of its high affinity to various tissues and its tendency to accumulates^{14,15,16} the result of the present study is inconsistent with other findings^{12,16,17}. The biological effects ascribed to Zingiber include induction of endogenous antioxidants in rat tissue organs, scavenged free radicals by giving electron(s) to them, protecting cells from oxidative stress and increased detoxification of foreign compounds¹⁸.

Serum Creatinine and urea are the most sensitive biochemical marker used in the assessment of renal tissue damage because the creatinine and urea are excreted through kidney therefore, in cellular damage there is retention of creatinine, urea in the blood¹⁹. Many previous studies reported that proteinuria from mercuric toxicity is due to disorders in the glomerular filtration barrier which resulted from damage in podocytes, and decreased reabsorption of filtered protein^{16,20}. The significant increase in serum creatinine level is evidence of the reduced ability of the renal tubules to excrete and creatinine from the blood of the HgCl₂ group rats, a reality that was also supported by the photomicrographs of the kidney tissue. Creatinine and urea levels

return to nearly normal levels in the (*Z officinale* + HgCl₂) group that give an indication that Zingiber has protective and ameliorative effects on HgCl₂ induced kidney damage (table 2). These results are inconsistent with a previous explanation that the zingiber contains a high concentration of flavonoids and alkaloids^{21,22} and acts as an antioxidant and/or free radical scavenging activity. Also the present study showed that treated of rats with the HgCl₂, markedly induced elevation in malondialdehyde (MDA) due to the induction of free

radical generation and stimulation of lipid peroxidation. Treatment of rats with Zingiber significantly reduces MDA level, due to Ginger compounds like gingerols, shagols, and ketone they have the capacity to reduce the free radical capacity^{22,23}.

The epithelium of (pct) is the most sensitive part of the kidney to the toxic effects of mercuric chloride due to their enzymatic activity^{4,24}. In a histomorphometric study the diameter of (pct) in the treated HgCl₂ group was significantly increased ($P \leq 0.05$) as compared with other experimental groups, this an agreement with previous results^{4,25,26}. One of the reasons for the increase in the diameter of (pct) is loss of brush border due to oxidative stress produced by HgCl₂^{21,22}. A significant decrease in the number of (pct) of the HgCl₂ reflects oxidative stress-induced necrosis and apoptosis^{27,28}. In the present study the maintenance of the diameter of (pct) in the (*Z officinale* + HgCl₂) group indicates the antioxidant and anti-inflammatory effects of Zingiber, as reported previously by^{26,29,30}.

Figure 2 shows that the HgCl₂ causes pathological changes and, On another side the morphological picture of *Z officinale* + HgCl₂ rat kidney group shows nearly normal tissue architecture with very few degenerative changes. These results were consistent with previously published articles^{4,24}. The result of the present study confirms the role of Zingiber in improving tissue damages by reducing/inhibit the number of pct and pathological changes induced by a toxic dose of HgCl₂.

Conclusion

Z officinale has an antioxidant effect and able to retard the progression of renal tubular pathological changes induced by HgCl₂.

Conflict of Interest Statement: The authors declare that there is no conflict of interest.

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References

1. Ghaleb A Oriquat, Tahia Saleem, Rajashri R Naik, Said Z Moussa. A Sub-Chronic Toxicity Study of Mercuric Chloride in the Rat. JJBS. 2012 Mar; 5(2)141-46.

2. Hui M, Wu Q, Wang S, Liang S, Zhang L, Wang F, et al. Mercury flows in China and global drivers. *Environ. Sci. Technol.* 2016 Nov; 51(1) 222–231.
3. Sheikh T J, Patel B J, Joshi D V, Patel R B. Repeated dose oral toxicity of inorganic mercury in Wistar rats: biochemical and morphological alterations. *Vetworld.* 2013 Jun; 6(8) 563-67.
4. Bridges C C, Zalups R K, Von Burg R. The aging kidney and the nephrotoxic effects of mercury. *J Toxicol Environ Health B Crit Rev.* 2017 Feb; 20(2) 55-80.
5. Orr SE, Bridges CC. Chronic kidney disease and exposure to nephrotoxic metals. *Int J Mol Sci.* 2017 May;18(5) 1039-1073.
6. Hosseini A, Rajabian A, Fanoudi S, Farzadnia M. Protective effect of Rheum turkestanicum root against mercuric chloride-induced hepatorenal toxicity in rats. *Avicenna J Phytomed.* 2018 Nov-Dec; 8(6) 488-97.
7. Wang WH, Wang ZM. Studies of commonly used traditional medicine-ginger. *China J. Chin. Mater. Med.* 2005 Oct; 30(20) 1569–73.
8. Khan RU, Naz S, Nikousefat Z, Tufarelli V, Javdani M, Qureshi M.S et al. Potential applications of ginger (*Zingiber officinale*) in poultry diets. *World's Poult. Sci. J.* 2012 Jun; 68(2) 245–52.
9. Miri Park, jungdon Bae, Dae-Sil Lee. Antibacterial activity of [10]-gingerol and [12]-gingerol isolated from ginger rhizome against periodontal bacteria. *Phytotherapy Research.* 2008 Nov; 22(11) 1446-49.
10. Bridges CC, Zalups RK. Transport of inorganic mercuric and methylmercuric in target tissues and organs. *J Toxicol Environ Health B Crit Rev.* 2010 Jun;13(5) 385-410.
11. Patel TA, Rao MV. Ameliorative effect of certain antioxidants against mercuric induced genotoxicity in peripheral blood lymphocytes. *Drug Chem Toxicol.* 2015 Feb; 38 (4) 408-14.
12. Dan Gao, Ling-Na Zeng, Pin Zhang, Zhi-Jie Ma, Rui-Sheng L, Yan-Ling Zhao, et al. Rhubarb Anthraquinones Protect Rats against Mercuric Chloride (HgCl₂)-Induced Acute Renal Failure. *Molecules.* 2016 Mar; 21(3) 298.
13. Radwanska K, Wazniak F, Siezieniewska, Z. "Influence of lead and mercury on the histological, ultrastructural and histochemical picture of the liver of albino rats." *Ann. Univ. Mariae. Cure Sklodowska Med.* 1993; 48(0)141-147.
14. Bernhoft RA. Mercury toxicity and treatment: a review of the literature. *J Environ Public Health.* 2011 Dec 22. doi: 10.1155/2012/460508;
15. Banerjee SK, Maulik M, Manchanda SC, Dinda AK. Garlic-induced alteration in rat liver and kidney morphology and associated changes in endogenous and oxidant status. *Food Chem Toxicol.* 2001 Aug; 39(8) 793-7.
16. Elshemy M M, Abdel-mejied A E-S, Zahran F, Omran. DPPD ameliorates renal fibrosis induced by HgCl₂ in rats. *Bioscience Research.* 2018 Sep; 15(3) 2416-25.
17. Aziza S, Elsenosi Y, Elsenosi A, Mogda K M. Role of antioxidant and anti-inflammatory of Ginger (*Zingiber officinal Roscoe*) against metalaxyl induced oxidative stress in rats. *Benha Vet Med J.* 2017 Dec; 33(2) 504-16.
18. Lopez-Giacoman S, Madero M. Salvador. Biomarkers in chronic kidney disease, from kidney function to kidney damage. *World J Nephrol.* 2015 Feb; 4(1): 57–73.
19. Boroushaki MT, Mollazadeh H, Rajabian A, Dolati K, Hoseini A, Paseban M, et al. Protective effect of pomegranate seed oil against mercuric chloride-induced nephrotoxicity in rat. *Renal Failure.* 2014 May; 36 (10):1581-6.
20. Hasan I H, EL-Desouky MA, Abdelaziz G M, Hozayen W G. Protective effects of *Zingiber officinale* against carbon tetrachloride-induced liver fibrosis. *International Journal of Pharmacy and Pharmaceutical Sciences.* 2016 Feb; 8(3) 377-81.
21. Salihu M, Ajayai B O, Adedara I A, Farombi E O. Gingerol-Rich Fraction from *Zingiber officinale* Prevents Hematotoxicity and Oxidative Damage in Kidney and Liver of Rats Exposed to Carbendazim. *J Diet Suppl.* 2016 Aug;13(4):433-48.
22. Tanko Y, Kabiru A, Abdulrazak A, Mohammed KA, Salisu AI, Jimoh A et al. Effects of Fermented Ginger Rhizome (*Zingiber officinale*) and Fenu Greek (*Trigonella foenum-graceum*) Supplements on Oxidative stress and Lipid Peroxidation Biomarkers in Poloxamer-407 Induced -Hyperlipidemic Wistar Rats. *Niger J Physiol Sci.* 2017 Dec;;32(2):137-43.
23. Kwanjit D, Jitprapa K, Bung-orn S, Suphat S. Antioxidant activity of ginger extract as a daily

- supplement in cancer patients receiving adjuvant chemotherapy: a pilot study. *Cancer Manag Res.* 2017 Jan; 9:11-18. doi: 10.2147/CMAR.S124016
24. Prabhu N S, Brijender B, Renu S. Evaluation of Protective Effect of *Tagetes erecta* Against Mercuric Chloride Induced Nephrotoxicity. *International Journal of Pharmaceutical and Clinical Research.* 2017 Aug; 9(8): 593-98.
25. Zaheer A, Talat Y, Irfan A, Khalida P. Lead-induced morphometric changes in the kidneys of albino rats ameliorated by Ginkgo biloba extract (EGb 761). *Journal of the Pakistan Medical Association.* 2017 Jan; 67(1): 58-65.
26. Chávez-Gómez N, Cabello-López A, Gopar-Nieto R, Aguilar-Madrid G, Aceves-Valdez M, et al. Enfermedad renal crónica en México y su relación con los metales pesados. *Rev Med Inst Mex Seguro Soc.* 2017 Nov-Dec; 55(6) 725-34.
27. Khan N, Naqvi A, Perveen K, Rafique M. Lead-induced nephrotoxicity with special reference to proximal tubule in albino rats. *Pak J Pharmacol.* 2008 Jan; 25(1)29-35.
28. Hamed MA, Ali SA, El-Rigal NS. Therapeutic potential of Ginger against renal injury induced by carbon tetrachloride in rats. *The Scientific World Journal.* 2012;2012 12 pages. <http://dx.doi.org/10.1100/2012/840421>
29. Stephane B, Charles R, Sylvain D, Yves C. The ginkgo biloba extract (EGb 761) protects hippocampal neurons against cell death induced by β -amyloid. *European Journal of Neuroscience.* 2000 Jun; 12(6)1882-90.
30. Morakinyo AO, Achema PU, Adegoke OA. Effect of *Zingiber officinale* (Ginger) on sodium arsenite-induced reproductive toxicity in male rats. *African Journal of Biomedical Research.* 2010Jan; 13(1) 39-45.