

Validation of a Simple Extraction Procedure for Stimulants Identification from Human Plasma

Idha Arfianti Wiraagni¹⁻⁴, Mustafa Ali Mohd², Rusdi Abdul Rashid³, Didi Erwandi bin Mohamad Haron²

¹Department of Pathology, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia, ²Shimadzu-UM Center for Xenobiotics Studies (SUCXeS), Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia,

³Department of Psychological Medicine, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia,

⁴Department of Forensic Medicine and Medicolegal, Faculty of Medicine, Universitas Gadjah Mada, Yogyakarta, Indonesia

Abstract

In this study, a novel LC-MS/MS (Liquid Chromatography-Tandem Mass Spectrometry) method was designed using a simple extraction procedure that was scientifically developed to capture the most relevant stimulants, methamphetamine, MDA (3,4-Methylenedioxyamphetamine), MDMA (3,4-Methylenedioxy-methamphetamine), and MDEA (Methyl diethanolamine). The LC-MS/MS method was validated using US FDA (United States Food and Drug Administration) guidelines, and all validation requirements were satisfactory. This protein precipitation method was specifically developed for handling large numbers of samples with minimum cost and volume of sample. The developed method was accurate, precise, and reproducible for quantification of stimulant from human plasma. This pilot study enrolled 100 drug addicts to evaluate stimulant testing from plasma. Our findings found 47 cases with positive methamphetamine. This method may be valuable for stimulant blood screening for a large and varied population because of its efficiency and economical aspects.

Key words : *Stimulant, LC-MS/MS, human plasma, protein precipitation*

Introduction

Stimulant covers many drugs including those that increase activity of the body and central nervous system, through various mechanism. Stimulant includes amphetamine, metamphetamine, caffeine, ephedrine, MDA, MDMA, MDPV, mephedrone, cocaine, nicotine, phenylpropanolamine, propylhexedrine, and pseudoephedrine. Metamphetamine, MDEA, MDA, and MDMA were studied in this research. Stimulants are used to treat sleep disorders, obesity, asthma, mood disorders, nasal congestion, impulse control disorders, and as anesthetics. Amphetamine colorless volatile liquid, has a burning taste and amine odor. It as being

very slightly soluble in water, but highly soluble in ethanol and chloroform, with a melting point of 25 °C, and boiling point 200-203°C.^{1,2}

Stimulants increase the presence neurotransmitter dopamine, noradrenaline, and serotonin. They usually block the reuptake or stimulate the efflux of neurotransmitter resulting in increased activity the nerve. Serotonin play a role in mood, appetite, thermoregulation, emotions, sleep, arousal, cognitive, autonomic functions, and pain regulatory systems. The elimination half-life is 6-12 hours. Both hepatic and renal are important organ to eliminate their metabolites from the body (Table 1).²

Corresponding author:

Idha Arfianti Wiraagni

E-mail: idha.arfianti@ugm.ac.id

<https://orcid.org/0000-0002-6532-0035> : Idha

Table 1: Pharmacokinetic profile of stimulants

No	Profile	MDMA/MDA/ MDEA	Amphetamine	Methamphetamine
1	Maximal concentration in the blood stream after ingestion	Between 1.5 and 3 hours.	Between 3 and 7 hours. Well absorbed from the gut.	Between 3.13 and 6.3 hours. Well absorbed from the gut.
2.	Target organ in distribution	Most part of the body, especially brain.		
3.	Duration of action	4-6 hours	3-6 hours	10-20 hours
4.	Metabolism	In liver use CYP450, CYP2D6, CYP3A4, and Catechol-O-methyltransferase	In liver use CYP2D6, glycine N-acyltransferase, dopamine β -hydroxylase, butyrate-CoA ligase, and flavin-containing monooxygenase 3	
5.	Elimination	Via kidney, about 20% is excreted unchanged in the urine.	Via kidney, about 30–40% of the drug unchanged in the urine.	Via kidney, about 30–54% of the drug unchanged in the urine.

Source : (2), (3)

Stimulant overdose symptoms and signs are rigid, jerking limbs, seizure, loss of coordination, clumsiness, muscle weakness, slurred or difficult speech, in and out consciousness, fast pulse, chest pain, irregular breathing, cognitive difficulties (confusion, poor judgment, fainting), and psychological distress (anxiety, paranoia, confusion, panicking, hallucinations or extremely agitated).⁴

Stimulants are frequently measured in urine or blood as screening for sports, employment, poisoning diagnostics, and medicolegal investigation with immunoassay or chromatography techniques (Table 2). The MDA concentration in the blood or urine of a person who taking MDMA is less than 10% of parent drug.³

Table 2: Analysis of stimulants in human biological sample using LC-MS/MS

No	Analyte	Sensitivity	Sample	Preparation	References
1	Stimulants	LOQ : 0.025-0.1 ng/mL	Urine	SPE	(5)
2	Stimulants	LOQ : 2.03–2.48 ng/mL	Plasma	Protein Precipitation	(6)
3	MDMA, MDA	LOQ : 100 ng/mL	Urine	SPE	(1)
4	Stimulants	LOQ : 2.5 ug/L	Blood, Urine	SPE	(7)

Materials and methods

Materials and standards

The Metamphetamine, MDEA, MDA, MDMA, and IS (Methamphetamine-d5, MDA-d5, and MDMA-d5) standard was purchased from Sigma-Aldrich (St. Louis, MO, USA). All solvents and reagents used were of HPLC grade and purchased from Merck (Darmstadt, Germany). Control plasma samples were obtained from the UMMC blood bank Malaysia.

Samples

A total of 100 blood samples were obtained from drug addiction that consenting to participation in the study. Demographic details such as gender, age, residential area, and ethnicity data were recorded. The inclusion criteria were 18 years of age or older, fully conscious, physically healthy, and no serious current psychiatric symptoms (i.e. psychotic episode). The exclusion criteria were refusal to participate, requiring advanced medical attention for a serious illness, and requiring psychiatric care for psychiatric symptoms. Institutional approval for the analysis of human samples was obtained from the Ethical Committee of University Malaya Medical Centre. The collection sample method used the method was published before.⁸

Extraction of plasma

Sample preparation was carried out using a protein precipitation extraction method. The frozen plasma was thawed at room temperature ($25 \pm 1^\circ\text{C}$). The thawed plasma was vortexed to ensure the sample was homogenous. To each 100 μL plasma sample, 50 μL of internal standard (IS) (containing 120 ng/mL of IS) was added, followed by the addition of 250 μL of acetonitrile (ACN). The mixture was vortexed, shaken for 5 s, and centrifuged for 2 min at 14800 rpm. The supernatant was filtered with a 0.2 μm syringe filter, then transferred to new a vial. Eight microliters were then injected into the LC-MS/MS system.

Liquid chromatography apparatus and conditions

In this research, the LC system consisted of an LC-20AD XR UFLC system with a SIL-HT automatic

sample injector (Shimadzu, Kyoto Japan) were used. The analytical column used was a Agilent, Eclipse Plus Phenyl-Hexyl (150 mm length x 2.1 mm ID, particle size 5 μm). Column temperature was 40 $^\circ\text{C}$ with total running time 10 min. Mobile phase used were 10 mM ammonium formate in water (pH 6.6) in pump A and 0.1% FA (in ACN) in pump B. The flow rate was set at 0.3 mL/min and a gradient elution was used at room temperature. The gradient program began with 5% B, then ramped to 40% B at 3.00 min, and held until 4.00 min. The gradient was ramped to 100% B at 6.00 min and held until 8.00 min. The gradient then returned to 5% B at 8.01 min and this condition was held for a further 10.00 min. Sample injection volume was 8 μL .

Mass spectrometry parameters

A Linear Ion Trap Quadrupole LC-MS/MS Spectrometer, QTRAP 5500, fitted with an ESI probe, and operated in the negative ionization mode was used to perform mass spectral analysis. The LC-MS/MS system was controlled by the Analyst software, version 1.6.3 (Applied Biosystems). Nitrogen was used as the nebulizer, auxiliary, collision, and curtain gas. Analytes were then quantified by multiple reactions monitoring (MRM). The optimal conditions were as follows: ion source temperature of 450 $^\circ\text{C}$, ion spray voltage of 5500 V, curtain gas of 20.0 psi, collision gas of medium, ion source gas 1 of 35.0 psi, and ion source gas 2 of 35.0 psi. Analyst 1.6.3 software was used for system control and data quantification.

Results

Linearity, sensitivity, and specificity

No significant interfering peaks were observed from the retention time corresponding to stimulants and the internal standards (Fig. 1-2). Seven calibration points were used to evaluate the linearity of the standard calibration curve. The standard curve was linear with 1/X as weighing factor and reached good linearity ($r > 0.997$). The signal to noise ratio which was greater than 10. The LOD for MDMA and methamphetamine was established at 0.25 ng/mL; MDA and MDEA was 1.25 ng/mL.

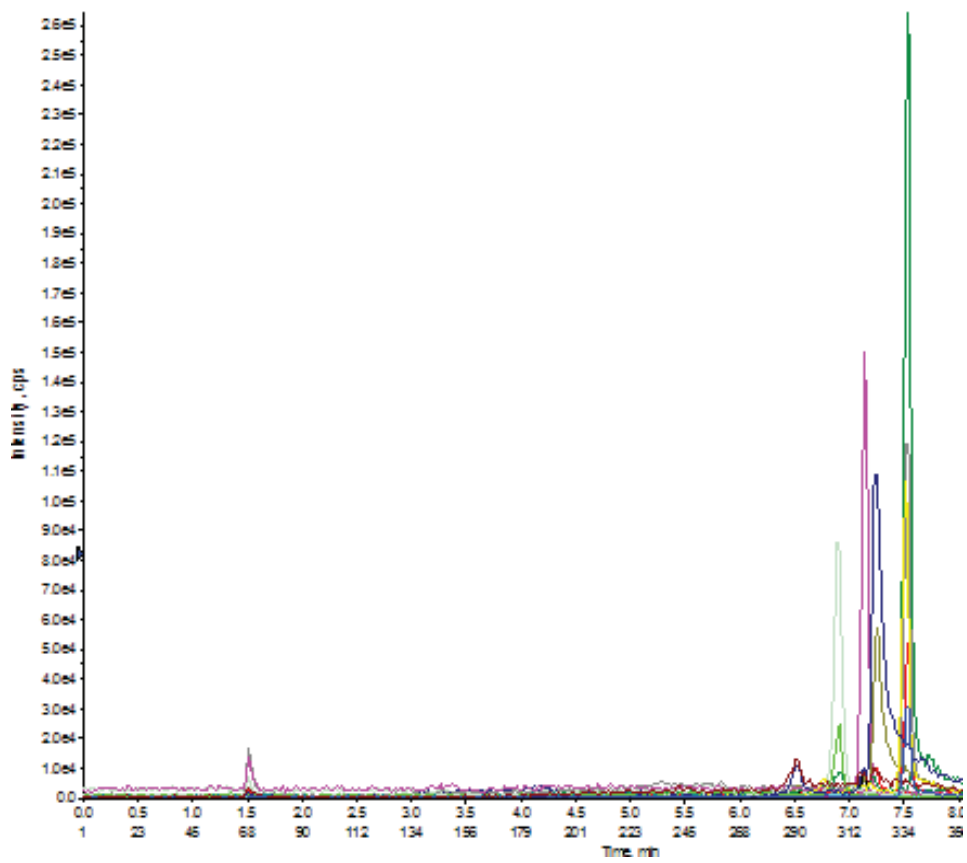


Fig. 1. Representative MRM chromatogram of blank.

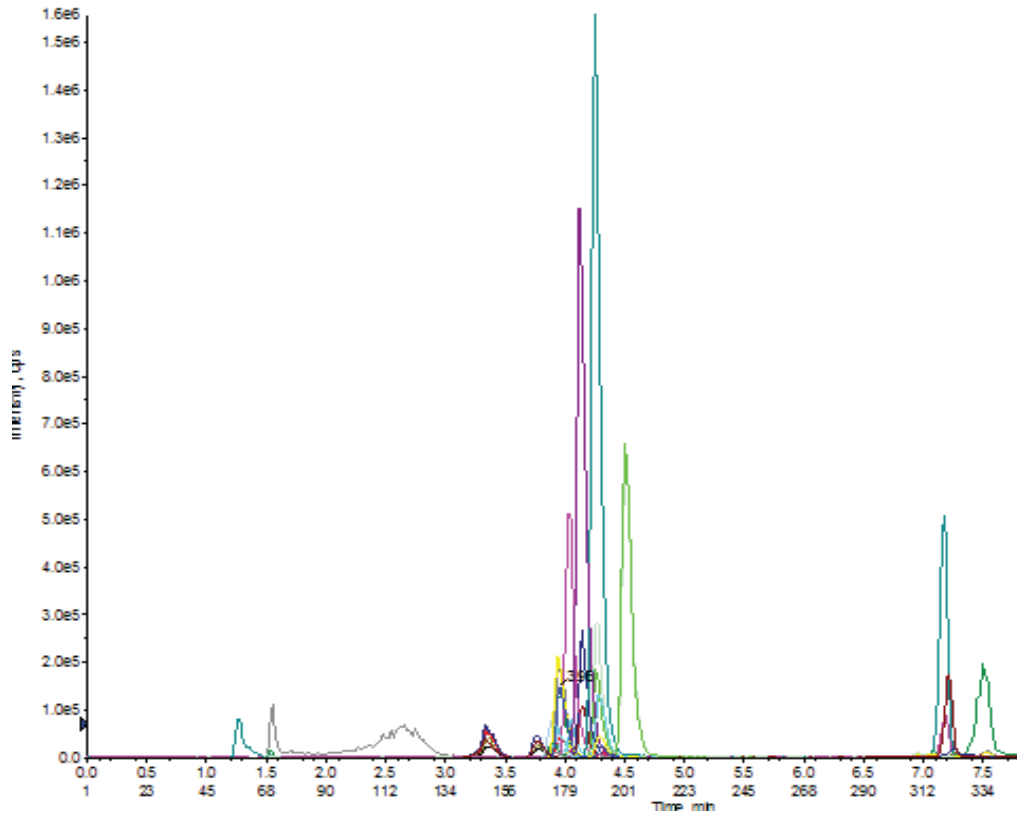


Fig. 2. Representative MRM chromatogram of stimulants in human plasma (QCL:1.5e6)

Precision and accuracy

Based on the mean percentage of coefficient of variation (%CV) for five quality control samples, the intra-day precision of plasma sample ranged from 0.41% to 9.93%. When assessed by means of the five quality control samples the accuracy for LLOQ intra-day plasma sample ranged from 98.16% to 118.4%, then for others ranged from 89.22% to 114%. The inter-day precision and accuracy were 7.17% to 9.97% and 95.4% to 113.12%. The results from this data clearly showed that the method that was developed has good accuracy, precision, and reproducibility for the quantification of stimulants from human plasma.

Recovery

The recovery of stimulants was tested at 0.5 and 5 ng/mL (Table 8). These results indicated that the extraction efficiency of stimulants using protein precipitation was quite good.

Table 3: Recoveries for DoA in human plasma

Compound	0.5 ppb	5 ppb
MDA	100%	91.9%
MDMA	97.5%	96.6%
Methamphetamine	96.8%	89.3%
MDEA	92.4%	88.1%

Stability testing

The stimulants precision ranged from 0.41% to 9.72% and accuracy ranged from 88.22% to 114% for freeze and thaw stability in plasma. The results indicated that the stimulants was stable in plasma for two cycles when stored at -20°C and thawed to room temperature. The precision and accuracy for stimulants bench top 4 hours stability in plasma ranged from 1.35% to 9.72% and from 95.08% to 114% respectively. The results indicated that the stimulants was stable in plasma at room temperature up to 4 hours when put on the bench. The precision and accuracy for stimulants autosampler stability in plasma ranged from 1.75% to 9.51% and from 98.54% to 114.2% respectively. The stimulants in plasma sample could be analyzed over 24 h in an autosampler at 20±1°C with satisfactory precision and accuracy. The precision and accuracy for stimulants long term (2 months) stability in plasma ranged from 1.59% to 8.87% and from 91.16% to 108.2% respectively.

Discussion

Analysis of plasma samples

This method was applied to the screening of stimulants levels in human blood plasma that were taken from 100 drug addicts of Malaysia (Table 4). Mostly participant from Kuala Lumpur and Selangor. Research subjects were 46.03±10.31 years old in average, with the youngest was 21 and the oldest was 72 years old. The mean of height and weight was 164.42±7.48 cm and 61.43±13.03 kg respectively.

Table 4. Baseline characteristics of participants.

Variable	N (%)	Variable	N (%)
Gender		Job Scope	
-Male	95 (95)	-Jobless	26(26)
-Female	5 (5)	-Private sector employee	74(74)
Age Group		Types of Food	
-18-30 yo	7 (7)	-Boiled	16 (16)
-31-40 yo	26 (26)	-Fried	84 (84)
-41-50 yo	34 (34)	-Raw	0 (0)
-51-60 yo	24 (24)	Source of Food	
-61 yo and above	9 (9)	-Home cooking	68 (68)
BMI		-Dining out	23 (23)

Table 4. Baseline characteristics of participants.

-Under weight	19 (19)	-Take away	9 (9)
-Normal weight	57 (57)	Smoking	
-Pre obesity	12 (12)	-Yes	85 (85)
-Obesity class 1	12 (12)	-No	15 (15)
Hometown		Source of Drinking	
-West Malaysia	149 (149)	-Treated water	72 (72)
-South Malaysia	1(1)	-Bottle water	28(28)
Race			
-Malaysian	80 (80)		
-Chinese	9 (9)		
-Indian	11 (11)		

In this study the majority of drug addicts were men. More addicts are men because they are braver and more easily influenced by the environment than women.. There are addicts who started taking drugs because of curiosity to try it. There are also those who use drugs to get recognition of their social status, also to feel the same experience with their friends. The results of this study are consistent with several other studies that show more male addicts than female. A study in Pakistan involving 150 respondents, was 100% male.^{9,10}

In this study most addicts were of working age. The possible cause is the heavy workload that exists and life style factor. Heavy workload, will be a heavy stressor too, so there are workers who are looking for pleasure by consuming drugs. According to combined 2008 to 2012 National Survey on Drug Use and Health (NSDUH), 113.1 million persons of US Population are full-time workers. In the other hand, some of the addicts don't even have a job. This is consistent with NSDUH, 13 million addicts in the USA do not work. There are some of the addicts who lost their jobs, because of using drugs.¹¹

Drug abusers tend to lose weight. This is because immunity in general decreases, so it often suffers from illness both mild illness and long-term pain, such as HIV and hepatitis. HIV and hepatitis are diseases that

can be transmitted through sharing needles and free sex between drug addicts. In 2007, the number of HIV in Russia among drug addicts reached 1.7 million, while the injecting drug user (IDU) with HIV reached 1.2 million. In 2002, the number of HIV in the USA among drug addicts reached 1.9 million, while the injecting drug user (IDU) with HIV reached 1.7 million. In 2005, the number of HIV in China among drug addicts reached 2.3 million, while the injecting drug user (IDU) with HIV reached 2.1 million.¹²

Chronic diseases will stimulate the body's immunity to be more active, and to stimulate metabolism faster. This can reduce the amount of fat in the body, if it continues continuously the addict will become underweight. In this study, the majority of addicts had a normal body mass index. The second place is underweight with some suffering from chronic diseases.¹³

Almost all addicts in this study were active smokers. This is consistent with studies in Pakistan, that the majority of addicts are active smokers. In drug addicts, the number of cigarettes smoked was affected by the type of drug consumed. Heroin users, had the highest cigarette consumption compared to other drugs.¹⁴ This can cause various respiratory illnesses. In addition, smoking will also put dozens of poisons into the body.¹⁰

From blood screening, we detected 47 cases with positive methamphetamine. Methamphetamine use, can be via oral, intranasal, smoking, and intravenous. The largest intravenous bioavailability, which reached 100%, while for smoking and oral about 67% and 79% intra nasal. Methamphetamine is metabolized in the liver by a process: N-demethylation to produce amphetamine, (ii) aromatic hydroxylation to produce primarily 4-hydroxymethamphetamine; and (iii) β -hydroxylation to produce norephedrine. Polymorphic cytochrome P450 2D6 causes different metabolisms in methamphetamine. Methamphetamine in the body has a different detection time, in the blood 24-48 hours, in saliva 24 hours, in urine up to 87 hours after taking 10 mg of methamphetamine tablets. Around 70% of a methamphetamine dose is excreted in the urine (within 24 hours) : 30-50% as methamphetamine, 10% as amphetamine, and up to 15% as 4-hydroxymethamphetamine. Acute effects will appear for more than 8 hours.¹⁵

Conclusions

This study successfully employed an inexpensive, rapid, and simple procedure for stimulants extraction and subsequent detection from human plasma by LC-MS/MS. The plasma extraction consisted of a simple protein precipitation method that consumes a low volume of organic solvent and trace volume of sample (100 μ l). The method was suitable for identifying and effectively separating stimulants with good precision and accuracy. This pilot study enrolled 100 drug addicts to evaluate stimulants testing from plasma. Our findings found 47 cases with positive methamphetamine. Determination of stimulants human plasma concentrations from the subjects could be satisfactorily performed by this proposed method.

Acknowledgements: We thank Shimadzu-UM Center for Xenobiotics Studies (SUCXeS) and the Pathology Department UM for laboratory service establishment in this study.

References

1. Nordgren HK, Beck O. Direct Screening of Urine for MDMA and MDA by Liquid Chromatography-Tandem Mass Spectrometry*. *Journal of Analytical Toxicology*. 2003;27: 15–19. doi:10.1093/jat/27.1.15
2. Lide DR, Baysinger G, Chemistry S, Berger LI, Goldberg RN, Kehiaian HV. *CRC Handbook of Chemistry and Physics*. : 2661.
3. Barnes AJ, De Martinis BS, Gorelick DA, Goodwin RS, Kolbrich EA, Huestis MA. Disposition of MDMA and Metabolites in Human Sweat Following Controlled MDMA Administration. *Clinical Chemistry*. 2009;55: 454–462. doi:10.1373/clinchem.2008.117093
4. Forray A, Sofuoglu M. Future pharmacological treatments for substance use disorders: Future pharmacotherapies for substance use disorders. *British Journal of Clinical Pharmacology*. 2014;77: 382–400. doi:10.1111/j.1365-2125.2012.04474.x
5. Feng J, Wang L, Dai I, Harmon T, Bernert JT. Simultaneous determination of multiple drugs of abuse and relevant metabolites in urine by LC-MS-MS. *Journal of analytical toxicology*. 2007;31: 359–368.
6. Vlase L, Popa D-S, Loghin F, Leucuta SE. High-throughput toxicological analysis of Methamphetamine, MDA and MDMA from human plasma by LC-MS/MS. *Romanian Journal of Legal Medicine*. 2009;17: 213–220.
7. Fernández M del MR, Wille SMR, Samyn N, Wood M, López-Rivadulla M, De Boeck G. High-Throughput Analysis of Amphetamines in Blood and Urine with Online Solid-Phase Extraction-Liquid Chromatography—Tandem Mass Spectrometry. *Journal of Analytical Toxicology*. 2009;33: 578–587. doi:10.1093/jat/33.9.578
8. Wiraagni IA, Mohd MA, Rashid R bin A, Haron DE bin M. Validation of a simple extraction procedure for bisphenol A identification from human plasma. *PLOS ONE*. 2019;14: 13.
9. Owczarek K, Kubica P, Kudlak B, Rutkowska A, Konieczna A, Rachoń D, et al. Determination of trace levels of eleven bisphenol A analogues in human blood serum by high performance liquid chromatography–tandem mass spectrometry. *Science of The Total Environment*. 2018;628–629: 1362–1368. doi:10.1016/j.scitotenv.2018.02.148
10. Khan MH, Anwar S, Khan IA, Khan RH, Subhan Z, Noreen N, et al. CHARACTERISTICS OF DRUG ABUSERS ADMITTED IN DRUG ABUSE TREATMENT CENTRES AT PESHAWAR, PAKISTAN. 2004;2: 4.
11. Kelly MG. Some Characteristics of Drug Abusers

- Attending a Drug Treatment Centre in Dublin. : 7.
12. Strathdee SA, Stockman JK. Epidemiology of HIV Among Injecting and Non-injecting Drug Users: Current Trends and Implications for Interventions. *Current HIV/AIDS Reports*. 2010;7: 99–106. doi:10.1007/s11904-010-0043-7
 13. Vc O. Demographic Characteristics as Predictors of Psychoactive Substance Use Dependence. *Addict Drug Abuse J* 2018, 1(1): 000104. 2018; 5.
 14. Stark MJ, Campbell BK. Drug use and cigarette smoking in applicants for drug abuse treatment. *J Subst Abuse*. 1993;5: 175–181.
 15. Cruickshank CC, Dyer KR. A review of the clinical pharmacology of methamphetamine. *Addiction*. 2009;104: 1085–1099. doi:10.1111/j.1360-0443.2009.02564.x