

Novel Mutations Associated with Amelogenesis Imperfecta- A Review

Vignesh P¹, Vijayashree Priyadharsini.J², Brundha M.P³

¹Research Associate, Dental Research Cell, ²Assistant Professor, Department of Microbiology, ³Associate professor, Department of Pathology, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, Chennai - TamilNadu, India

Abstract

Amelogenesis imperfecta [AI] is a congenital disorder that presents with a rare abnormal formation of the enamel, external layer of the crown of teeth, unrelated to any systemic or generalized Conditions. AI causes teeth to be usually small, discolored, pitted or grooved and prone to rapid wear and breakage. These defects which vary among affected individuals can affect both primary teeth and secondary teeth. About 14 forms of amelogenesis imperfecta have been described so far. They are distinguished by their specific dental abnormalities and by their pattern of inheritance. Additionally, amelogenesis imperfecta can occur alone without any other signs and symptoms or as a part of a syndrome that affects multiple parts of the body.

Keywords: *Amelogenesis imperfecta; gene mutation; enamel; dental.*

Introduction

Amelogenesis imperfecta [AI] is a congenital disorder that presents with a rare abnormal formation of the enamel, external layer of the crown of teeth, unrelated to any systemic or generalized conditions.^{1,2} Tooth enamel is the most highly mineralized structure in the human body, with 85.1% of its volume involved by hydroxyapatite crystals³. The physical properties and physiological function of enamel are directly related to the orientation, disposition Composition, and morphology of the mineral components within the tissue⁴. During organogenesis, the enamel transitions from a pliable tissue, which is almost entirely devoid of proteins.⁵ During genesis deflection of the unique molecular and cellular activities might be responsible for the patterns observed in amelogenesis imperfecta.⁶

The exact incidence of amelogenesis imperfecta is uncertain. Estimates vary wildly, from 1 in 700 people in northern Sweden to 1 in 14,000 people in the United states⁷, among which hypoplastic AI represents 60 - 73% of all cases, hypomaturation AI represents 20-40%, and hypocalcification AI represents 7%⁸. A complex interplay of genes results in the formation of enamel, growth and enamel mineralization. The most severe type is mostly caused by truncating mutations in the *FAN83H* gene^{9,10}.

Classification of Amelogenesis Imperfecta

Amelogenesis imperfecta can be classified based on the phenotype¹¹, based on the clinical, microradiography and histopathological findings, and on the phenotype and mode of inheritance^{12,13}. Most commonly accepted classification is the one being proposed by Witkop in the year 1988, which classified AI into mainly 4 types *viz.*, hypoplastic AI, hypomaturation AI and hypocalcified type based on developmental stages of enamel and hypoplastic-hypomaturation with taurodontism with several other subtypes under each type¹⁴.

Corresponding Author:

Vijayashree Priyadharsini.J

Assistant Professor, Department of Microbiology
Saveetha Dental College, Saveetha Institute of
Medical and Technical Sciences (SIMATS), Saveetha
University, Chennai - TamilNadu, India - 600077
E-mail: vijayashreej.sdc@saveetha.com

Table 1: Different types of amelogenesis imperfecta

Type 1 - Hypoplastic	IA hypoplastic, pitted autosomal dominant IB hypoplastic, local autosomal dominant IC hypoplastic, local autosomal recessive ID hypoplastic, smooth autosomal dominant IE hypoplastic, smooth X-linked dominant IF hypoplastic, rough autosomal dominant IG enamel agenesis, autosomal recessive
Type 2 - Hypomaturation	IIA hypomaturation pigmented autosomal recessive IIB hypomaturation IIC snow capped teeth, X-linked IID autosomal dominant
Type 3 - Hypocalcified	IIA autosomal dominant IIB autosomal recessive
Type 4 - Hypomature hypoplastic enamel with taurodontism	IVA hypomaturation - hypoplastic with taurodontism, autosomal dominant IVB hypoplastic - hypomaturation with taurodontism, autosomal dominant

1. Type 1 - Hypoplastic

The enamel of abnormal thickness due to malfunction in an enamel matrix formation. Enamel is extremely thin but hard & translucent and should have random pits & grooves. Condition is of an autosomal dominant, autosomal recessive, or x-linked pattern. Enamel differs in appearance from dentine radiographically as normal functional enamel.^{15,16}

2. Type 2 - Hypomaturation

Enamel has a sound thickness, with a pitted appearance. It is less hard compared to normal enamel, and is prone to rapid wear, although not as intense as Type 3 AI.¹⁷ Condition is of an autosomal dominant, autosomal recessive, or x-linked pattern. Enamel appears to be comparable to dentine in its radiodensity on radiographs.¹⁸

3. Type 3 - Hypocalcified

Enamel defect because of malfunction of enamel calcification, therefore enamel is of normal thickness but is extremely brittle, with an opaque/chalky presentation. Teeth are susceptible to staining and rapid wear, exposing dentin. Condition is of an autosomal dominant and autosomal recessive pattern. Enamel appears less radiopaque compared to dentine on radiographs.²⁰

4. Type 4 - Hypomature hypoplastic enamel and taurodontism

Enamel has a variation in appearance, with mixed features from Type 1 and Type 2 AI. All Type 4 AI has taurodontism in common. Condition is of the autosomal dominant pattern. Other common features may include an anterior open bite,¹⁹ taurodontism, the sensitivity of teeth. Differential diagnosis would include

dental fluorosis, molar-incisor hypomineralization, chronological disorders of tooth development.²⁰

INHERITANCE PATTERNS OF AI

AI may be inherited in three manners which are X-linked form, autosomal dominant, or autosomal recessive trait. Apparently molecular genetic tools will allow more precise diagnosis.^{21,22} Different genes are related to different forms of inheritance, located at different genomic sites. *AMELX* is associated with X-linked form which displays disorganized hypoplastic enamel.²³ Another gene shows both autosomal dominant and autosomal recessive inheritance patterns within the 4q13.3. has been identified as being allocated with on autosomal recessive inheritance.²⁴

1. X-linked forms of amelogenesis imperfecta

X-linked amelogenesis imperfecta (XAI) shows the typical pattern of X-linked inheritance. Heterozygous females can transfer the mutant gene to children of either sex with the danger of this being 50% The condition affects males and females in strikingly different ways. Males show the trait fully²⁵. They may have teeth that have only a thin layer of enamel of normal color and translucency, or the enamel may be of normal thickness but poorly mineralized with loss of translucency and/or a yellow-brown discoloration²⁶

2. Autosomal dominant amelogenesis imperfecta

Autosomal dominant AI (ADAI) typically affects one or more individuals in each generation of a family.²⁷ There may be consistency in the clinical manifestations in every affected individual or there may be the variable expression, resulting in substantial or subtle differences between different affected individuals in the same family²⁸. The phenotype in ADAI could also be predominantly or exclusively hypoplastic, manifested by thin enamel and spacing between the teeth, or in some pedigrees by rough, irregular, or randomly pitted enamel.^{29,30} If the prime defect is in the amount of enamel matrix produced, the enamel will be hard, normally translucent, and not subject to significant attrition.³¹

3. Autosomal recessive amelogenesis imperfecta

Autosomal recessive AI (ARAI) should be considered if there is known consanguinity in a family

with an affected individual. This may be more often encountered in certain ethnic and cultural groups where intermarriage within the family may be more common (for example, AI in association with cone rod dystrophy, a syndromic condition^{32,33}. ARAI will also be more prevalent where there is a high frequency of the mutant gene in a population, such as in some Polynesian communities³⁴

NOVEL MUTATION ASSOCIATED WITH AI

The first AI causing mutation was identified in the gene encoding the enamel matrix protein known to make up the bulk of the secreted enamel organic matrix³²[TABLE 2]. The EMP gene evolved from a standard ancestral gene and formed a part of the secretory calcium-binding phosphate gene cluster. The enamel matrix protein include amelogenin [*AMELX*] which makes up around 90%, of the EMD,³³ secrete *AMELX* last with remaining ameloblastin (AMBN) and *ENAM*³⁴

GENE	Mode of inheritance
FAM83H	Autosomal dominant
FAM20A	Autosomal recessive
AMELX	X-linked form
ENAM	Autosomal dominant
MMP20	Autosomal recessive
KLK-4	Autosomal recessive
WDR72	Autosomal recessive
SLC4A4	Autosomal recessive
LAMB3	Autosomal dominant
ITGB6	Autosomal recessive

Table 2: Different gene mutation reported in AI

Enamel is a highly mineralized structure in the body with 85% of its volume occupied by hydroxyapatite crystals.³⁵ During the organogenesis, the enamel metamorphoses to form soft, pliable time to its final form which is almost devoid of protein. The final composition of enamel is a reflection of its unique molecular and cellular activity that happens during its genesis³⁶ deviation from this pattern may lead to amelogenesis imperfecta (AI)³⁷. With an underlying genetic cause, the etiology of AI includes febrile illness or vitamin deficiency, local infection or trauma, fluoride ingestion congenital syphilis, birth defects or idiopathic factors³⁸. *ENAM* and *AMELX* encode extracellular matrix proteins of the developing enamel and *KLK-4* and *MMP20* encode proteases that help corrupt organic matter from the enamel matrix during the development phase of amelogenesis³⁹. *SLC24A4* encodes a calcium transporter that intervenes calcium transport to develop enamel during tooth development. Less is thought about the function of other genes implicated in amelogenesis imperfecta.⁴⁰

MANAGEMENT OF AI

The management of individuals affected by AI has been described in three phases viz., temporary phase, transcriptional phase and permanent phase. The first phase of management is directed during primary and mixed dentition.³⁵ The transcriptional phase occurs during the period of tooth eruption and continues till adulthood and the final stage or permanent phase occurs in adulthood. In primary dentition, the dental treatment of affected children aims to ensure favorable conditions for the eruption of the permanent teeth as well as for the normal growth of the facial bones and TMJ.³⁶ In the mixed dentition, the treatment goals are to preserve tooth structure, maintain Tooth Vitality, decrease tooth sensitivity, vertical dimension, and Improve esthetics. when permanent first molars and anterior teeth erupt orthodontic and prosthetic assessment is essential.³⁷ However, rehabilitation in the mixed dentition is complex, since teeth have different eruption sequence, and definitive treatment cannot be rendered until the complete eruption of permanent dentition.³⁸ In the permanent dentition, the final treatment objective is to diminish tooth sensitivity and to restore the vertical dimension of occlusion, the function of well as esthetic is the final treatment often starts is soon in the clinical

height of the crown and gingival tissue have been stabilizes and the pulp time has reduced.³⁹

Conclusion

Amelogenesis imperfecta is a hereditary disorder affecting enamel often leading to physiological stress in patients resulting in poor aesthetics. One of the greatest challenges faced by the clinician in the total rehabilitation.⁴⁰⁻⁴² The treatment plan with respect to AI patients may be influenced by factors such as age, socioeconomic status and other comorbid conditions. Early plans for treatment, precise surgical procedure in order to meet the aesthetic and functional demands of patients with modern technology is warranted in the present scenario.^{43,44}

Acknowledgement: We thank Saveetha Dental College for providing us the support to conduct the study

Conflict of Interest:

The authors declared that there is no conflict of interest.

Source Of Funding: Self

Ethical Clearance: Not Required

References

1. Slootweg P. *Pathology of the Maxillofacial Bones: A Guide to Diagnosis*. Springer, 2015.
2. Kida M, Ariga T, Shirakawa T, et al. Autosomal-dominant hypoplastic form of amelogenesis imperfecta caused by an enamelin gene mutation at the exon-intron boundary. *J Dent Res* 2002; 81: 738-742.
3. Robinson C, Briggs HD, Atkinson PJ, et al. Matrix and mineral changes in developing enamel. *J Dent Res* 1979; 58: 871-882.
4. Mahoney EK, Rohanizadeh R, Ismail FSM, et al. Mechanical properties and microstructure of hypomineralised enamel of permanent teeth. *Biomaterials* 2004; 25: 5091-5100.
5. Paine ML, White SN, Luo W, et al. Regulated gene expression dictates enamel structure and tooth function. *Matrix Biol* 2001; 20: 273-292.
6. Simmer JP, Fincham AG. Molecular mechanisms of dental enamel formation. *Crit Rev Oral Biol Med*

- 1995; 6: 84–108.
7. Rajendran R. Chapter 1. Developmental disturbances of oral and paraoral structures. *Shafer's Textbook of Oral Pathology, 5th Edition, Elsevier, Amsterdam*; 67.
 8. Renuka S, Np M. COMPARISON IN BENEFITS OF HERBAL MOUTHWASHES WITH CHLORHEXIDINE MOUTHWASH: A REVIEW. *Asian Journal of Pharmaceutical and Clinical Research* 2017; 3–7.
 9. Xin W, Wenjun W, Man Q, et al. Novel FAM83H mutations in patients with amelogenesis imperfecta. *Sci Rep* 2017; 7: 6075.
 10. Witkop CJ. Amelogenesis imperfecta, dentinogenesis imperfecta and dentin dysplasia revisited: problems in classification. *Journal of Oral Pathology and Medicine* 1988; 17: 547–553.
 11. Weinmann JP, Svoboda JF, Woods RW. Hereditary disturbances of enamel formation and calcification. *The Journal of the American*, [https://jada.ada.org/article/S0002-8177\(45\)27001-X/abstract](https://jada.ada.org/article/S0002-8177(45)27001-X/abstract) (1945).
 12. WITKOP, J C. Heritable defects of enamel. *Oral Facial Genetics* 1976; 151–226.
 13. Bäckman B, Holm AK. Amelogenesis imperfecta: prevalence and incidence in a northern Swedish county. *Community Dent Oral Epidemiol* 1986; 14: 43–47.
 14. Stephanopoulos G, Garefalaki M-E, Lyroudia K. Genes and related proteins involved in amelogenesis imperfecta. *J Dent Res* 2005; 84: 1117–1126.
 15. Seow WK. Clinical diagnosis and management strategies of amelogenesis imperfecta variants. *Pediatr Dent* 1993; 15: 384–393.
 16. Nevelle BW, Damm DD, Allen CM, et al. Oral and Maxillofacial Pathology, (2dedn). *Saunders New Delhi India*.
 17. Marickar RF, Geetha RV, Neelakantan P. Efficacy of contemporary and novel Intracanal medicaments against enterococcus faecalis. *J Clin Pediatr Dent* 2014; 39: 47–50.
 18. Herzog CR, Reid BM, Seymen F, et al. Hypomaturation amelogenesis imperfecta caused by a novel SLC24A4 mutation. *Oral Surg Oral Med Oral Pathol Oral Radiol* 2015; 119: e77–81.
 19. Bouvier D, Duprez JP, Bois D. Rehabilitation of young patients with amelogenesis imperfecta: a report of two cases. *ASDC J Dent Child* 1996; 63: 443–447.
 20. Seow WK, Lai PY. Association of taurodontism with hypodontia: a controlled study. *Pediatr Dent* 1989; 11: 214–219.
 21. Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. An insight into the emergence of *Acinetobacter baumannii* as an oro-dental pathogen and its drug resistance gene profile - An in silico approach. *Heliyon* 2018; 4: e01051.
 22. Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. In silico analysis of virulence genes in an emerging dental pathogen *A. baumannii* and related species. *Arch Oral Biol* 2018; 94: 93–98.
 23. Gibson CW, Yuan ZA, Hall B, et al. Amelogenin-deficient mice display an amelogenesis imperfecta phenotype. *J Biol Chem* 2001; 276: 31871–31875.
 24. Hart TC, Hart PS, Gorry MC, et al. Novel ENAM mutation responsible for autosomal recessive amelogenesis imperfecta and localised enamel defects. *J Med Genet* 2003; 40: 900–906.
 25. Paine ML, Lei Y-P, Dickerson K, et al. Altered Amelogenin Self-assembly Based on Mutations Observed in Human X-linked Amelogenesis Imperfecta (AIH1). *Journal of Biological Chemistry* 2002; 277: 17112–17116.
 26. Pratha AA, Geetha RV. Awareness on Hepatitis-B vaccination among dental students-A Questionnaire Survey. *J Adv Pharm Technol Res* 2017; 10: 1360.
 27. Sohaib Shahzan M, Smiline Girija AS, Vijayashree Priyadharsini J. A computational study targeting the mutated L321F of ERG11 gene in *C. albicans*, associated with fluconazole resistance with bioactive compounds from *Acacia nilotica*. *J Mycol Med* 2019; 29: 303–309.
 28. Girija As S, Priyadharsini J V. CLSI based antibiogram profile and the detection of MDR and XDR strains of *Acinetobacter baumannii* isolated from urine samples. *Med J Islam Repub Iran* 2019; 33: 3.
 29. Smiline A, Vijayashree JP, Paramasivam A. Molecular characterization of plasmid-encoded

- blaTEM, blaSHV and blaCTX-M among extended spectrum β -lactamases [ESBLs] producing *Acinetobacter baumannii*. *Br J Biomed Sci* 2018; 75: 200–202.
30. Paramasivam A, Vijayashree Priyadharsini J, Raghunandhakumar S. N6-adenosine methylation (m6A): a promising new molecular target in hypertension and cardiovascular diseases. *Hypertens Res* 2020; 43: 153–154.
 31. Kim J-W, Seymen F, Lin BP-J, et al. ENAM mutations in autosomal-dominant amelogenesis imperfecta. *J Dent Res* 2005; 84: 278–282.
 32. O'Hara MC, Guatelli-Steinberg D. Differences in enamel defect expression and enamel growth variables in *Macaca fascicularis* and *Trachypithecus cristatus* from Sabah, Borneo. *Journal of Archaeological Science* 2020; 114: 105078.
 33. Smillie AC, Rodda JC, Kawasaki K. Some aspects of hereditary defects of dental enamel, including some observations on pigmented Polynesian enamel. *N Z Dent J* 1986; 82: 122–125.
 34. Girija SA, Jayaseelan VP, Arumugam P. Prevalence of VIM- and GIM-producing *Acinetobacter baumannii* from patients with severe urinary tract infection. *Acta Microbiol Immunol Hung* 2018; 65: 539–550.
 35. Gisler V, Enkling N, Zix J, et al. A multidisciplinary approach to the functional and esthetic rehabilitation of amelogenesis imperfecta and open bite deformity: a case report. *J Esthet Restor Dent* 2010; 22: 282–293.
 36. Ranta H, Lukinmaa PL, Waltimo J. Heritable dentin defects: nosology, pathology, and treatment. *Am J Med Genet* 1993; 45: 193–200.
 37. Geetha RV, Veeraraghavan VP. Evaluation of antibacterial activity of five root canal sealants against enterococcus faecalis-an in vitro study. *Int J Pharm Sci Rev Res* 2016; 40: 221–223.
 38. Gibbard PD. The management of children and adolescents suffering from amelogenesis imperfecta and dentinogenesis imperfecta. *Int J Orthod* 1974; 12: 15–25.
 39. Kurian M, Geetha RV. Effect of herbal and fluoride toothpaste on *Streptococcus mutans*-a comparative study. *Res J Pharm Biol Chem Sci* 2015; 7: 864.
 40. Ashwin KS, Muralidharan NP. Vancomycin-resistant enterococcus (VRE) vs Methicillin-resistant *Staphylococcus Aureus* (MRSA). *Indian J Med Microbiol* 2015; 33 Suppl: 166–167.
 41. Girija ASS, Smiline Girija AS, Vijayashree Priyadharsini J, et al. Plasmid-encoded resistance to trimethoprim/sulfamethoxazole mediated by *dfrA1*, *dfrA5*, *sul1* and *sul2* among *Acinetobacter baumannii* isolated from urine samples of patients with severe urinary tract infection. *Journal of Global Antimicrobial Resistance* 2019; 17: 145–146.
 42. Shahana RY, Muralidharan NP. Efficacy of mouth rinse in maintaining oral health of patients attending orthodontic clinics. *Research Journal of Pharmacy and Technology* 2016; 9: 1991–1993.
 43. Vaishali M, Geetha RV. Antibacterial activity of Orange peel oil on *Streptococcus mutans* and *Enterococcus-An* In-vitro study. *Research Journal of Pharmacy and Technology* 2018; 11: 513–514.
 44. Geetha RV, Thangavelu L, Others. Evaluation of anti-inflammatory action of *Laurus nobilis*-an in vitro studyf anti-inflammatory action of *Laurus nobilis*-an in vitro study. *International Journal of Research in Pharmaceutical Sciences* 2019; 10: 1209–1213.