

# Association of *Retn* (*Rs3219175*) Polymorphism with *Osc* Progression and Development - An *In Silico* Study

Monica.K<sup>1</sup>, J. Vijayashree Priyadharsini<sup>2</sup>, Gheena. S<sup>3</sup>, Pratibha Ramani<sup>4</sup>, Abhilasha. R<sup>5</sup>,

<sup>1</sup>Research Associate, Dental Research Cell, <sup>2</sup>Assistant Professor, Department of Microbiology,

<sup>3</sup>Associate Professor, Department of Oral Pathology and Microbiology, <sup>4</sup>Professor and Head of the Department, Department Of Oral Pathology and Microbiology, <sup>5</sup>Reader, Department of Oral Pathology and Microbiology,

<sup>6</sup>Senior Lecturer, Department of Oral Pathology and Microbiology, Saveetha Dental College and Hospitals, Saveetha Institute of Medical And Technical Science, Saveetha University, Chennai

## Abstract

Oral squamous cell carcinoma (OSCC) is more common with 90% of malignant neoplasm in oral cavity. Multiple factors leading to OSCC like genetic alterations, chronic exposure, micro environmental carcinogens, premalignant disease like oral epithelial dysplasia, mitochondrial membrane damage and viruses infections like HPV (human papillomavirus). *RETN* gene belongs to the family of mouse resistin-like genes. Human genetic variations are found to influence inter-individual variation observed in disease susceptibilities, drug response, response to an infectious disease. At the molecular level these sequence variations influence gene expression, protein production, function, disease susceptibility in particular individuals. The present *in silico* study determines the possible association of *RETN* polymorphism with that of oral squamous cell carcinoma in different populations. In this study the *RETN* gene polymorphism *rs321975* was analysed in comparison to the global frequencies. The present study attempts to identify the possible reason for the selection of "A" allele in the south asian population. Furthermore, genotyping analysis has to be performed in a south Indian population to ascertain the association between *RETN* polymorphism (*rs3219175*) and OSCC.

**Keywords:** Polymorphism; oral squamous cell carcinoma; *RETN*; variant

## Introduction

Oral squamous cell carcinoma (OSCC) is more common, which accounts for 90% of malignant neoplasm in oral cavity. Worldwide, oral cancer accounts for 2% - 4% of all cancer cases, tongue as a more common subtype<sup>1,2</sup>. In some regions, the prevalence of oral cancer is higher, 45% in India & 10% in Pakistan<sup>3,4</sup>. The field cancerization has been explained as in altered telomeres and unbalanced allelic loci seen in both tumors and

surrounding histologically normal tissues at distances of least 1 cm from the visible tumor margins results in the occurrence of multiple primary cancers in the head and neck region and recurrence following complete excision of the original tumor<sup>5,6</sup>. Multiple factors rising OSCC which are genetic alterations, chronic exposure, micro environmental carcinogens<sup>7</sup>, premalignant disease to oral cancer can occur with alteration seen in the stromal tissue<sup>8</sup>. Oral epithelial dysplasia also has a malignant transformation potential<sup>1</sup>, mitochondrial membrane damage leads to apoptosis<sup>9,10</sup> and viruses like HPV (human papillomavirus) also plays an important role in oral cancer<sup>11</sup>. Numerous risk factors associated with osc are chronic inflammation, alcohol and tobacco consumption, betel quid chewing and viral infections<sup>12</sup>. It is known that tumour cells on course of development undergo molecular alterations in cellular molecules including DNA, RNA, and proteins which could be attributed to the inherent biological properties of the

---

### Corresponding Author:

**Vijayashree Priyadharsini**

Assistant Professor, Department of Microbiology,  
Saveetha Dental College And Hospitals,  
Saveetha Institute of Medical And Technical Science,  
Saveetha university, Chennai.

E mail id: vijayshreej.sdc@saveetha.com

cancer cell<sup>13,14,15</sup>. The survival rate has reduced due to its diagnosis in the advanced stage<sup>16</sup>. There is a need for molecular prognostic factors to detect patients with high risk of oral cancer recurrence and treatment failure. The upregulation genes were seen involved in extracellular matrix degradation and epithelial mesenchymal transition, downregulation seen in detoxification pathways<sup>17,18</sup>. Expression of multinucleated giant cells shows host immune response<sup>19,20,15</sup>.

Latest reports from the International Agency for Research on Cancer (IARC) with ICD-10 code = C06 : lip, oral cavity in 2018 reports shows incidence around the world is 3,54,864 and in India it is 1,19,992. Prevalence of 9,13,514 reported around the world and In India it is around 2,65,225. Mortality rate reported around the world is 1,77,384 and in India around 72,616 cases. The *RETN* gene belongs to the family of mouse resistin-like genes. Hormone released suppresses insulin ability which stimulates glucose uptake in adipose cells. Studies have shown links between obesity and diabetes. It also helps promote chemotaxis in myeloid cells - gene Cards<sup>12,21</sup>. Single nucleotide polymorphisms (SNP) is common DNA sequence variation which influences gene expression, protein production, function, disease susceptibility in high risk individuals<sup>7,12,21,22</sup>. Resistin levels of plasma correlates with the inflammatory markers and coronary artery calcification, and plays an important role in the measure of coronary atherosclerosis<sup>22-24</sup>. Kumar et al, studied genetic variations of *RETN* associated with greater risk with various diseases like metabolic syndrome and colon cancer<sup>22,23</sup>. A study reported that upregulation of resistin gene expression in human breast cancer tissues<sup>25</sup>. The present *in silico* study determines the selection of *RETN* SNPs and its possible association with oral squamous cell carcinoma.

## Materials and Methods

One of the regulatory region variants of the *RETN* (gene was selected for the study based on the literature mining process. The minor allele frequency was found to be 0.08 (A allele). Since oral cancer is a disease that is more predominant in patients with a chronic history of using smoking and smokeless tobacco use such as pan, gutka etc., genetic variants which were more closely related to environmental and habitual factors were identified in the gene and selected. As most of

these factors are known to affect the inflammatory pathways leading to disease a study carried out by Yang et al was selected for comparison. The authors declared that the AA homozygous state of *RETN* polymorphism (*rs3219175*) presented a high risk of advanced tumor size when compared to GG homozygous genotype. The Ensembl database was used to acquire the frequency data of the variant in different populations. The data presented by Yang et al., was selected as a reference population (Taiwanese), which was used for further comparisons. Gene expression analysis would provide clues on the putative role of *RETN* gene in HNSCC. Hence, gene expression profile for different grades of tumor was analysed using the UALCAN database in the HNSC dataset. Survival curve analysis based on the Kaplan-Meier method was also investigated for differential expression of *RETN* within the patient group.<sup>26,27</sup>

## Results and Discussion

The phenotype database from ensembl database for *rs3219175* polymorphism showed 92% of "G" and 8% of "A" allele polymorphism reported, represented in figure 1. Individual allele frequencies of different populations are represented in figure 2. Analysis of allele frequency among different population like African (G=87% , A=13%), East Asian (G=82% , A=18%), South Asian (G=98% , A=2%) and European (G=99% , A=1%) was compared with global population (G=92% , A=8%). Allele frequency which is observed in the East Asian population showed 82% of "G" allele and 18% of "A" allele within different sub-populations such as Chinese Dai in Xishuangbanna, China(CDX), Han Chinese in Beijing, China (CHB), Southern Han Chinese(CHS), Japanese in Tokyo, Japan(JPT), Kinh in Ho Chi Minh City, Vietnam (KHV). Allele frequency observation in South-Asians population showed 98% of "G" allele frequency and 2% of "A" allele frequency which is compared among the sub-population of BEB (Bengali from bangladesh), GIH (Gujarathi Indians from Houston), ITU (Indians Telugu from UK), PJI (Punjabi from Lahore), STU ( Sri Lankan Tamil from UK) among which "A" allele is higher in BEB and "G" allele is high in STU population (data not shown). To derive a more accurate picture on the deviations observed for the polymorphism in different populations, we compared the allele frequency from the Yang et al., (Taiwanese

population) study of the data acquired from the Ensembl database. The allele frequency comparison was almost similar (Figure 3 and 4). But when comparing the frequency of alleles between the south Asian and the ancestral population which is the African population, we observed marked differences in the allele frequency. The “A” allele frequency was higher in African and east Asian population when compared to other groups. Furthermore, the A allele was negatively selected in other populations such as American, south Asian and European. A negative selection of the allele in the population may be indicative of the fact that it could be deleterious in some way, but the reason for such deviation has to be analysed to derive a strong conclusion.

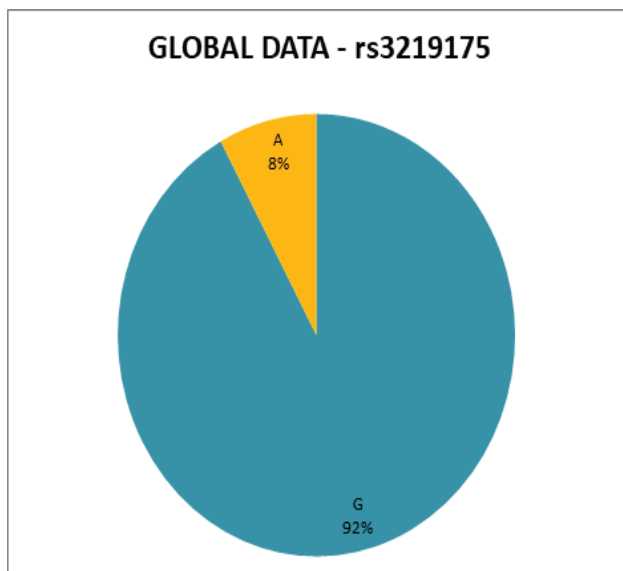


Figure 1: The global phenotype data from Ensembl database for *rs3219175* polymorphism (Yellow color denotes “A” allele frequency = 8%; Blue color denotes “G” allele frequency = 92%)

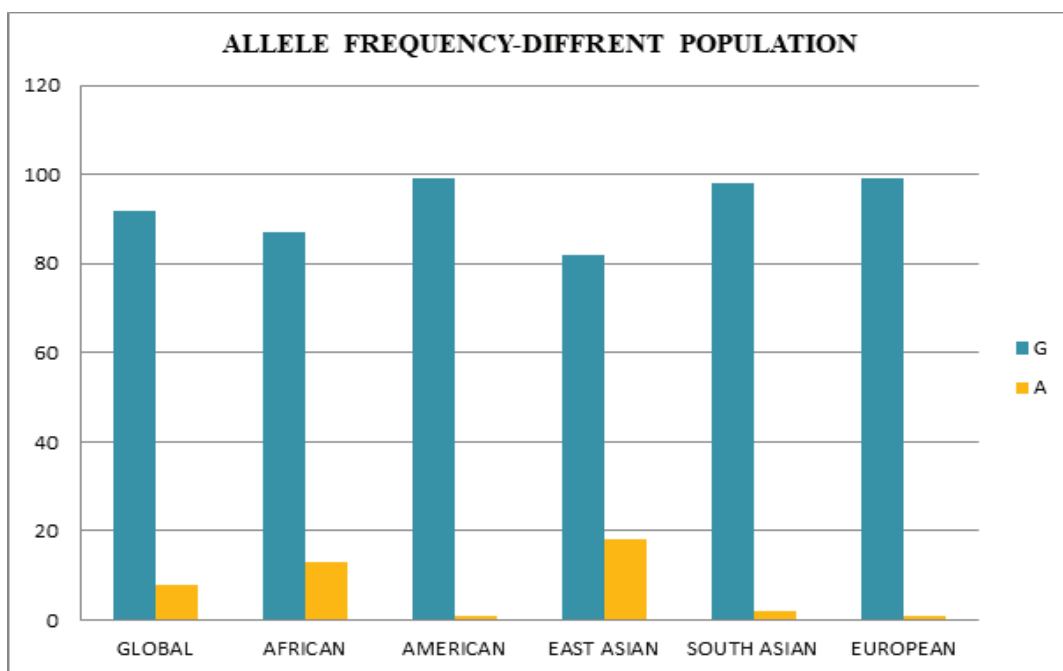


Figure 2: Analysis of allele frequency for *rs3219175* polymorphism among different populations. Yellow color denotes “A” allele frequency; Blue color denotes “G” allele frequency. [Global G - 92% and A- 8%; African - G-87% and A-13%; American -G-99% and A-1%; East Asian - G-82% and A-18%; European - G-99% and A-1%; South Asian - G-98% and A-2%].

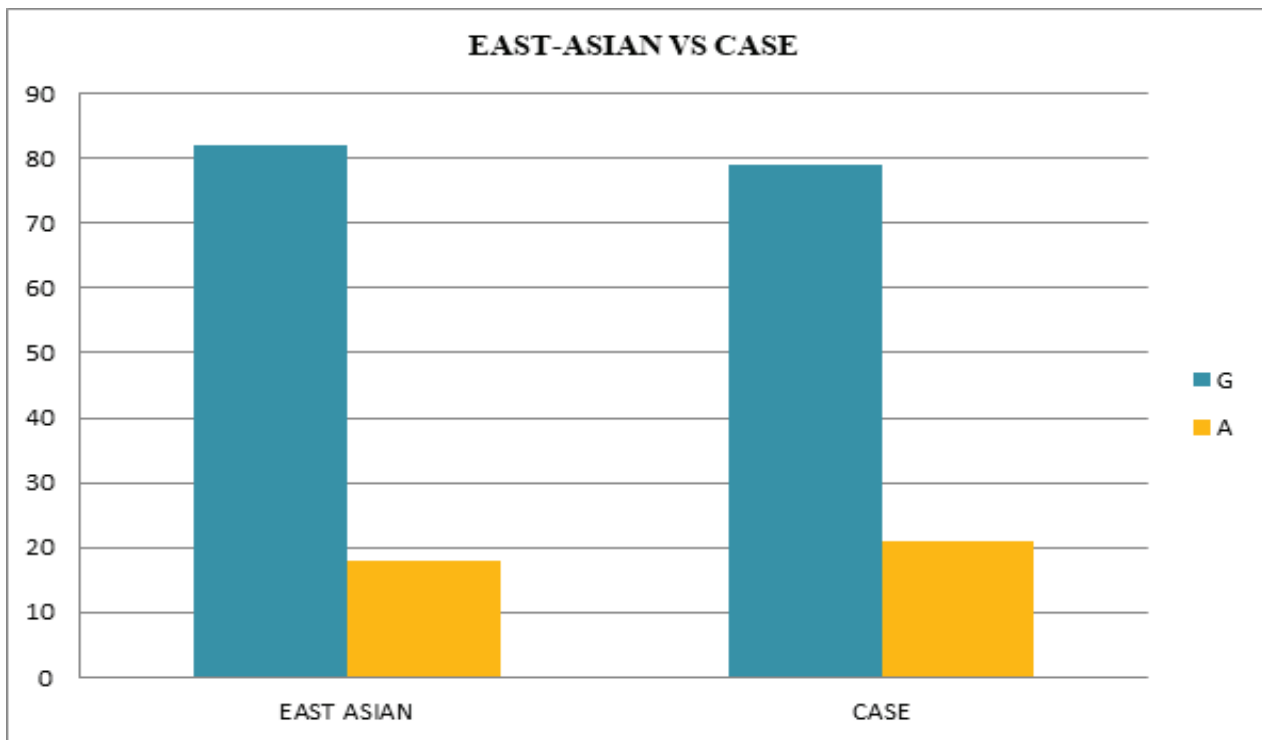


Figure 3: Comparison of *rs3219175* allele frequency case data from research study by Yang et al (G = 78% and A = 22%) with East Asian population (G = 82% and A = 12%). Yellow color denotes “A” allele frequency; Blue color denotes “G” allele frequency.

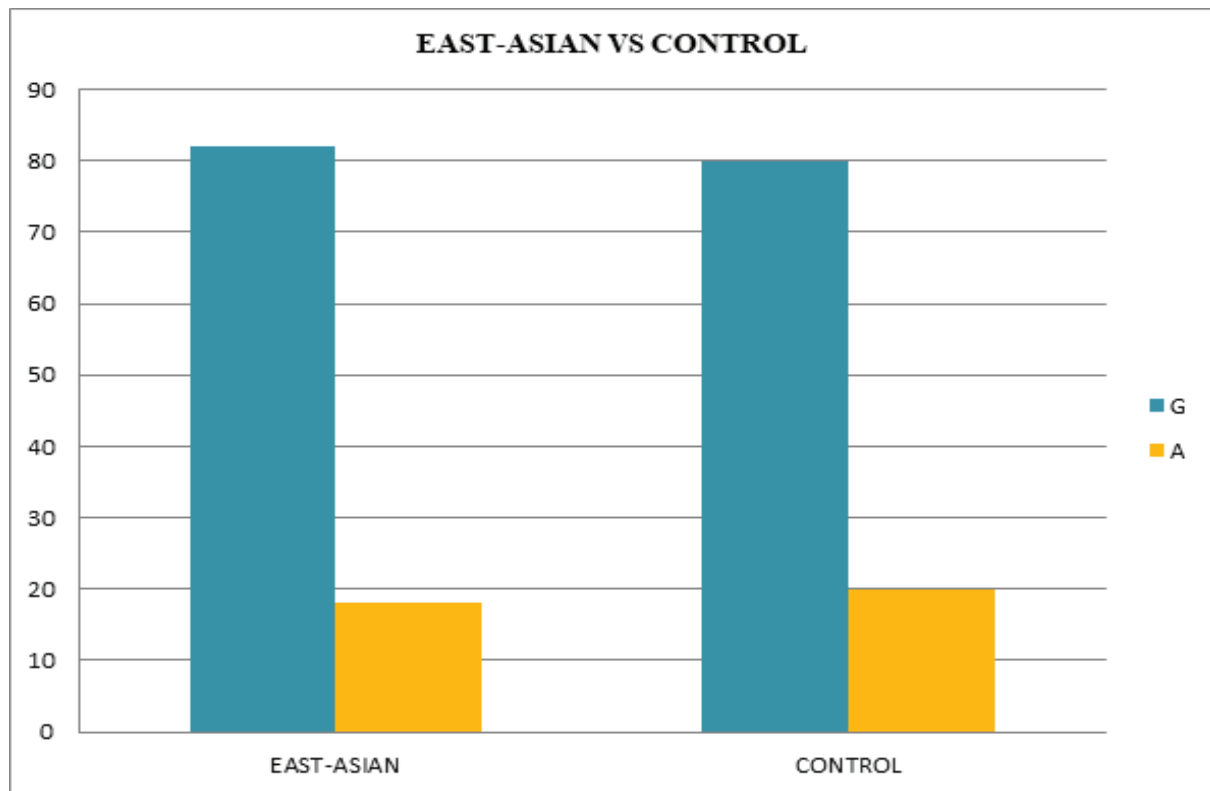
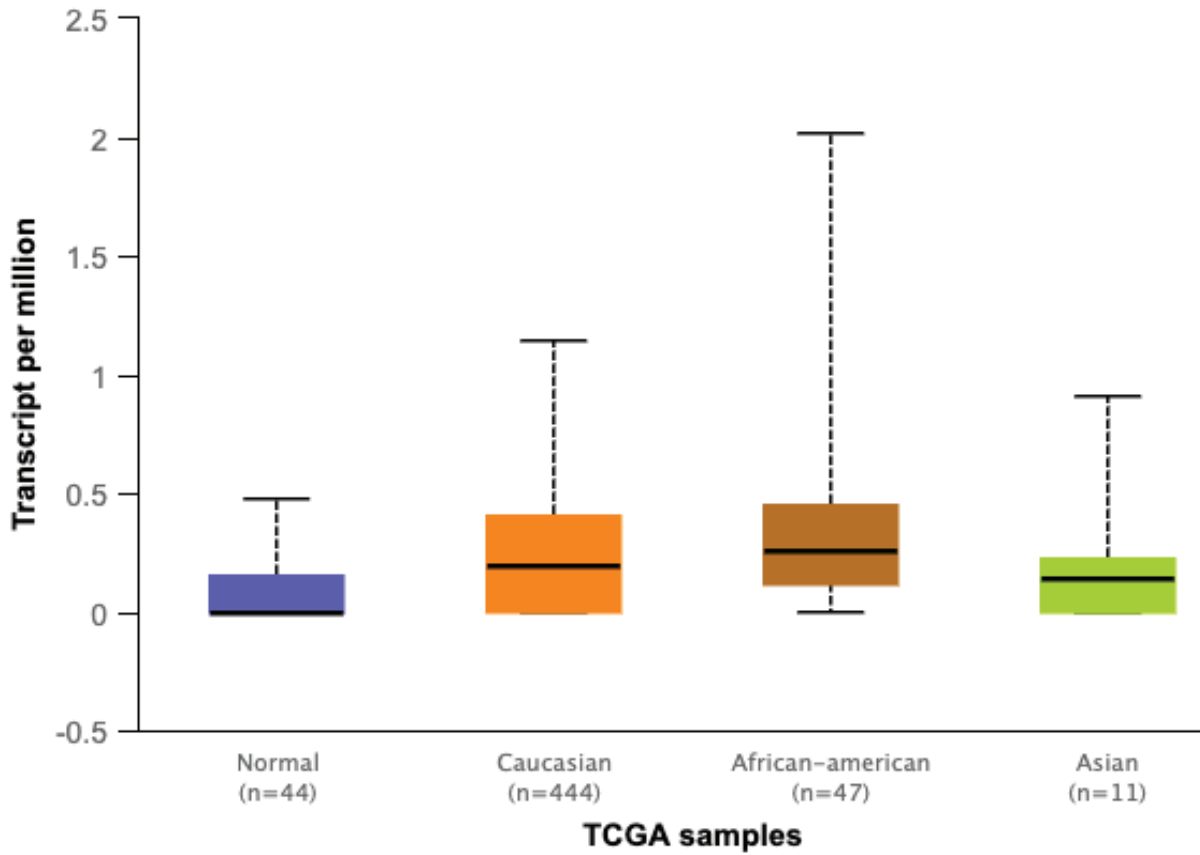
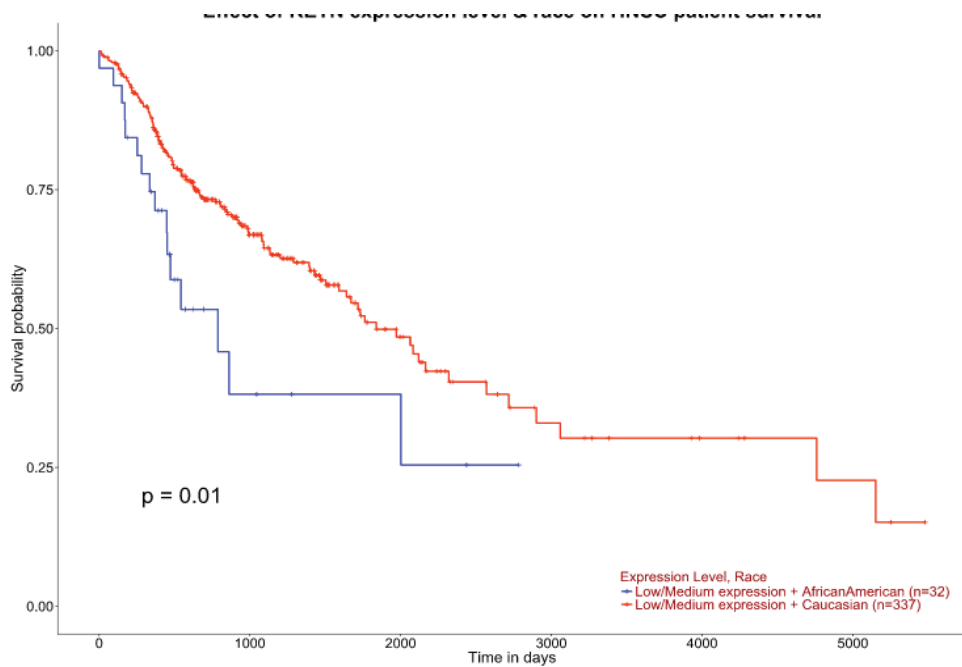


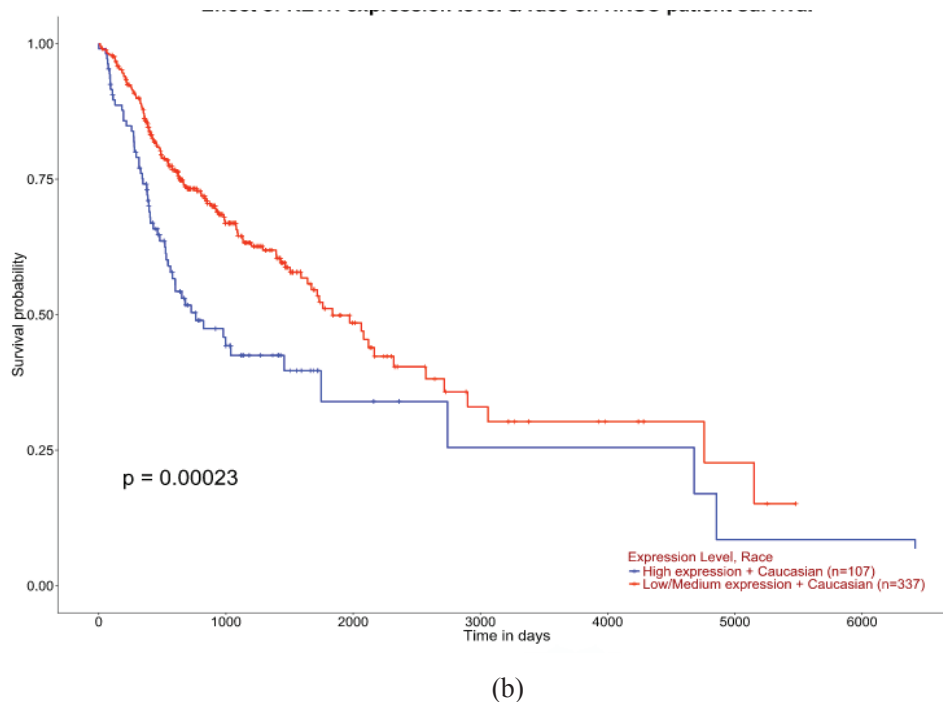
Figure 4: Comparison of *rs3219175* allele frequency of control data (G = 80% and A = 20%) from the research study by Yang et al with East Asian population (G = 82% and A = 12%). Yellow color denotes “A” allele frequency; Blue color denotes “G” allele frequency.



**Figure 5: Expression of *RETN* in HNSC based on the patient’s race. Blue color denotes normal population; Orange color denotes Caucasian population; brown color denotes African-American population; green color denotes Asian population. The comparison of gene expression between different races returned significant values between the group African-American vs Asian ( $p=4.4 \times 10^{-02}$ ).**



(a)



**Figure 6: (a) Comparison of low/medium *RETN* gene expression between African American and Caucasian patients produced a significant p value of 0.01; (b) Comparison of high and low/medium *RETN* gene expression between Caucasian patients produced a significant p value of <0.01.**

In addition, expression of the *RETN* gene found to differ significantly between the African American population and Asian population (Figure 5). When analysing the gene expression pattern between groups, low/medium level expression between African American and Caucasians returned a significant p value of 0.01, wherein low/medium gene expression in African Americans corroborated with poor survival of HNSC patients. Interestingly, we identified a significant difference between high and low/medium expression in Caucasian patients, where high level expression resulted in poor survival of these patients (Figure 6). These results provided preliminary evidence on the pivotal role of the *RETN* gene in the progression of HNSC. It is a known fact that nature tries to eliminate anything that is not in use<sup>28,9</sup>. Resistin is an adipokine that is associated with obesity, inflammation and various study cancer.<sup>7</sup> Resistin levels may play a role in pathogenesis of cancer cachexia<sup>7,12</sup>. Upregulation of serum Resistin has been detected in OSCC patients<sup>7</sup>. Online databases are an excellent source for identifying differential gene expression, which is a popular source for exploration of gene expression in diseases. The modern biomedical investigators can genetically profile diseases with

this organised online database. There are various tools with basic designs available which are needed in experimenting *in silico* studies<sup>7,29</sup>. The “A” allele frequency is dramatically increased in the ancestral and east Asian population which creates an inquisitiveness to analyze the gene and the polymorphism to draw an association between the gene and the disease phenotype. The negative selection of “A” allele especially in the south Asian population is of our interest.

## Conclusion

The present *in silico* study has provided preliminary data regarding the population wide-analysis of the *RETN* polymorphism selected. The deviation in allele frequency clearly indicates that the gene is evolving and is under constant process of natural selection. As a thumb rule, the alleles which are known to be deleterious are negatively selected, whereas a beneficial allele is positively selected in a population. The observation here denotes that the A allele could be a deleterious allele, and may confer risk of tumor progression in OSCC patients. Further experimental evidence are warranted to provide firm evidence on the association of *RETN* gene mutations or variations with that of OSCC.

**Acknowledgement:** The authors would like to acknowledge the help and support rendered by the department of oral pathology and the information technology and management of saveetha dental college for the constant assistance with the research.

**Conflicts of Interest:** The authors declare no potential conflict of interest.

**Source of Funding:** Self

**Ethical Clearance:** It is taken from “Saveetha Institute Human Ethical Committee” (Ethical Approval Number- SDC/SIHEC/2020/DIASDATA/0619-0320)

### References:

- Jayaraj G, Ramani P, Herald J, Sherlin, Premkumar P, Anuja N. Inter-observer agreement in grading oral epithelial dysplasia – A systematic review [Internet]. Vol. 27, Journal of Oral and Maxillofacial Surgery, Medicine, and Pathology. 2015. p. 112–6. Available from: <http://dx.doi.org/10.1016/j.ajoms.2014.01.006>
- Viveka TS, Shyamsundar V, Krishnamurthy A, Ramani P, Ramshankar V. p53 Expression Helps Identify High Risk Oral Tongue Premalignant Lesions and Correlates with Patterns of Invasive Tumour Front and Tumour Depth in Oral Tongue Squamous Cell Carcinoma Cases [Internet]. Vol. 17, Asian Pacific Journal of Cancer Prevention. 2016. p. 189–95. Available from: <http://dx.doi.org/10.7314/apjcp.2016.17.1.189>
- Williams HK. Molecular pathogenesis of oral squamous carcinoma. Mol Pathol. 2000 Aug;53(4):165–72.
- Siddiqui IA, Farooq MU, Siddiqui RA, Rafi SMT. Role of toluidine blue in early detection of oral cancer. Pak J Med Sci Q. 2006;22(2):184.
- Gupta V, Ramani P. Histologic and immunohistochemical evaluation of mirror image biopsies in oral squamous cell carcinoma [Internet]. Vol. 6, Journal of Oral Biology and Craniofacial Research. 2016. p. 194–7. Available from: <http://dx.doi.org/10.1016/j.jobcr.2016.06.002>
- Hashibe M, Boffetta P, Janout V, Zaridze D, Shangina O, Mates D, et al. Esophageal cancer in Central and Eastern Europe: Tobacco and alcohol [Internet]. Vol. 120, International Journal of Cancer. 2007. p. 1518–22. Available from: <http://dx.doi.org/10.1002/ijc.22507>
- Yang W-H, Wang S-J, Chang Y-S, Su C-M, Yang S-F, Tang C-H. Association of Resistin Gene Polymorphisms with Oral Squamous Cell Carcinoma Progression and Development. Biomed Res Int. 2018 Oct 14;2018:9531315.
- Jayaraj G, Sherlin HJ, Ramani P, Premkumar P, Natesan A. Stromal myofibroblasts in oral squamous cell carcinoma and potentially malignant disorders. Indian J Cancer. 2015 Jan;52(1):87–92.
- Hannah R, Ramani P, Sherlin HJ, Ranjith G, Ramasubramanian A, Jayaraj G, et al. Awareness about the use, Ethics and Scope of Dental Photography among Undergraduate Dental Students Dentist Behind the lens. J Adv Pharm Technol Res. 2018;11(3):1012.
- Gheena S, Ezhilarasan D. Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells. Hum Exp Toxicol. 2019 Jun;38(6):694–702.
- Jayaraj G, Sherlin HJ, Ramani P, Premkumar P, Anuja N. Cytomegalovirus and Mucoepidermoid carcinoma: A possible causal relationship? A pilot study. J Oral Maxillofac Pathol. 2015 Sep;19(3):319–24.
- Husain N, Neyaz A. Human papillomavirus associated head and neck squamous cell carcinoma: Controversies and new concepts. J Oral Biol Craniofac Res. 2017 Sep;7(3):198–205.
- Sethi S, Ali S, Philip PA, Sarkar FH. Clinical advances in molecular biomarkers for cancer diagnosis and therapy. Int J Mol Sci. 2013 Jul 16;14(7):14771–84.
- Sridharan G, Ramani P, Patankar S. Serum metabolomics in oral leukoplakia and oral squamous cell carcinoma. J Cancer Res Ther. 2017 Jul;13(3):556–61.
- Swathy S, Gheena S, Varsha SL. Prevalence of pulp stones in patients with history of cardiac diseases [Internet]. Vol. 8, Research Journal of Pharmacy and Technology. 2015. p. 1625. Available from: <http://dx.doi.org/10.5958/0974-360x.2015.00291.7>
- Hema Shree K, Ramani P, Sherlin H, Sukumaran G, Jeyaraj G, Don KR, et al. Saliva as a Diagnostic Tool in Oral Squamous Cell Carcinoma - a Systematic Review with Meta Analysis. Pathol Oncol Res. 2019 Apr;25(2):447–53.
- Thangaraj SV, Shyamsundar V, Krishnamurthy

- A, Ramani P, Ganesan K, Muthuswami M, et al. Molecular Portrait of Oral Tongue Squamous Cell Carcinoma Shown by Integrative Meta-Analysis of Expression Profiles with Validations. *PLoS One*. 2016 Jun 9;11(6):e0156582.
18. Jangid K, Alexander AJ, Jayakumar ND, Varghese S, Ramani P. Ankyloglossia with cleft lip: A rare case report. *J Indian Soc Periodontol*. 2015 Nov;19(6):690–3.
  19. Kumar A, Sherlin HJ, Ramani P, Natesan A, Premkumar P. Expression of CD 68, CD 45 and human leukocyte antigen-DR in central and peripheral giant cell granuloma, giant cell tumor of long bones, and tuberculous granuloma: An immunohistochemical study. *Indian J Dent Res*. 2015 May;26(3):295–303.
  20. Sridharan G, Ramani P, Patankar S, Vijayaraghavan R. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma. *J Oral Pathol Med*. 2019 Apr;48(4):299–306.
  21. Institute NC, National Cancer Institute. RETN Gene [Internet]. Definitions. 2020. Available from: <http://dx.doi.org/10.32388/lizinj>
  22. Shastri BS. SNP alleles in human disease and evolution [Internet]. Vol. 47, *Journal of Human Genetics*. 2002. p. 0561–6. Available from: <http://dx.doi.org/10.1007/s100380200086>
  23. Kumar S, Gupta V, Srivastava N, Gupta V, Mishra S, Mishra S, et al. Resistin 420C/G gene polymorphism on circulating resistin, metabolic risk factors and insulin resistance in adult women [Internet]. Vol. 162, *Immunology Letters*. 2014. p. 287–91. Available from: <http://dx.doi.org/10.1016/j.imlet.2014.07.009>
  24. [No title] [Internet]. [cited 2020 Jun 6]. Available from: <https://doi.org/10.1161/01.CIR.0000155620.10387.43>
  25. Vallega KA, Liu N, Myers JS, Yu K, Sang Q-XA. Elevated Resistin Gene Expression in African American Estrogen and Progesterone Receptor Negative Breast Cancer [Internet]. Vol. 11, *PLOS ONE*. 2016. p. e0157741. Available from: <http://dx.doi.org/10.1371/journal.pone.0157741>
  26. Website [Internet]. [cited 2020 Jun 11]. Available from: Hunt SE, McLaren W, Gil L, Thormann A, Schuilenburg H, Sheppard D, et al. Ensembl variation resources. Database [Internet]. 2018 Jan 1 [cited 2020 Jun 10];2018. Available from: <https://academic.oup.com/database/article-abstract/doi/10.1093/database/bay119/5255129>
  27. Chandrashekar DS, Bashel B, Balasubramanya SAH, Creighton CJ, Ponce-Rodriguez I, Balabhadrapatruni V S, et al. UALCAN: A Portal for Facilitating Tumor Subgroup Gene Expression and Survival Analyses [Internet]. Vol. 19, *Neoplasia*. 2017. p. 649–58. Available from: <http://dx.doi.org/10.1016/j.neo.2017.05.002>
  28. Sivaramakrishnan SM, Ramani P. Study on the Prevalence of Eruption Status of Third Molars in South Indian Population [Internet]. Vol. 07, *Biology and Medicine*. 2015. Available from: <http://dx.doi.org/10.4172/0974-8369.1000245>
  29. Murray D, Doran P, MacMathuna P, Moss AC. In silico gene expression analysis – an overview. *Mol Cancer*. 2007 Aug 7;6(1):1–10.