

Overview of the Content of Bisphenol A in the Amniotic Fluid of Pregnant Women And Its Adverse Health Outcomes

Dardan Dreshaj¹, Flaka Pasha²

¹Researcher, University of Prishtina "Hasan Prishtina", Kosovo, ²Researcher, Department of Pharmacology and Toxicology and Clinical Pharmacology, University of Prishtina "Hasan Prishtina", Kosovo

Abstract

Background: This review represents a critical and constructive analysis of literature in the content of Bisphenol A in amniotic fluid of pregnant women and its adverse health outcomes. The review is generated through summary, classification, analysis and comparison of already existing material and researches on field.

Methods : Databases as Scopus, PubMed, Medline, Web of Science, Global Health, were used to extract data for the review. Search terms like "Bisphenol A", "pregnancy", "amniotic fluid", "endocrine disruptors" were used. Out of 200 research articles screened, 70 most relevant studies are included in this review.

Conclusions: This review highlights the correlation between Bisphenol A and its endocrine disrupting potential, impacting especially fragile categories as pregnant women and their fetuses through its toxicokinetic features and its metabolites. Thus, BPA distorts important physiological processes necessary for fetal development and disease progression later in life.

We can conclude that human exposure to BPA, as one of the leading environmental contaminants, represents a major global issue and its adverse health outcomes can be debilitating for human health. Further research on field considering BPA distribution, varying exposure rates, racial disparities, inter-species differences and EDCs cumulative effects, should be conducted. Finding safer alternatives for replacing BPA in market must remain a priority.

Keywords: amniotic fluid; Bisphenol A, pregnancy; adverse health outcomes;

Introduction

Bisphenol A (BPA), 2,2-bis(4-hydroxyphenyl) propane (CAS No. 80-05-7), is a synthetic chemical excessively produced worldwide.¹ BPA commercial production began in the United States in 1957, followed by Europe one year later. Its global production grows consistently, varying between 0% and 5% annually, with the strongest growth occurring in China.² A yearly increase of 4.6% in production of BPA is envisioned to happen from 2013 to 2019.³

Bisphenol A is used to manufacture polycarbonate plastics and epoxy resins.⁴ Out of the total production of BPA, 65% is dedicated to polycarbonate synthesis, 25% for epoxy resins and the rest 10% for our daily use products such as food storage containers, food antioxidants, metal food cans, baby bottles, thermal papers, dental seals, medical devices, personal care products, safety helmets, sunglasses, lenses, cosmetics, infant incubators, CD/DVDs, hair dryers, fridges, computers and smartphones.⁵

Bisphenol A is synthesized by the condensation of phenol with acetone in the presence of a catalyst, a strongly acidic ion-exchange resin. BPA consists of a central tetrahedral carbon atom with two methyl and two phenol groups. BPA is relatively water-soluble (120-

Correspondence:

Dr. Flaka Pasha,

Str: Fehmi Agani 141, Prishtine, 10000, Kosovo

Email: flaka.pasha@uni-pr.edu; +383 44584005

300 mg/L),⁶ dissociates in alkaline environment, has a moderate bioaccumulation rate, low vapor pressure, high melting point, and low half-life in air (0.2 days).^{7,8}

BPA leaches into environment and impacts human health during its production, processing or waste disposal.⁹ BPA leaching also occurs when polycarbonate and epoxy resin-containing products are exposed to heat, are re-used or their pH changes.¹⁰ BPA products in contact with heat, acidic or basic conditions accelerate the hydrolysis of the ester bonds between BPA molecules, thus exposing humans to its harmful metabolites. This happens when people heat cans to sterilize food, place acidic or basic food in cans or polycarbonate plastic, and keep heating or washing these products.^{11,12} Even though ingestion of contaminated food is the most common way how human get exposed to BPA, inhalation and skin absorption are considered to be of great importance too.^{13,14}

Also, moderate levels of active unconjugated BPA have been detected in human serum, adipose tissue, breast milk, placenta, maternal and fetal plasma, indicating that BPA can accumulate in human body.^{15,16,17}

The United States Environmental Protection Agency (EPA) reported that more than 400,000 kilograms of BPA leach into environment every year¹⁸, alarming us on the great burden of exposure, BPA's cumulative effects and life-long disadvantageous health impact.

Bisphenol A mechanisms of action and its disrupting potential

BPA is qualified as a xeno-estrogen that disturbs synthesis, transport, activity and metabolism of endogenous estrogens, consequently affecting the development, growth and reproduction of organisms.^{19,20}

BPA can mimic or antagonize endogenous hormones, subsequently perturbing endocrine function, by binding weakly to several steroid receptors including the estrogen receptors (ER α and β) and thyroid hormone receptor.^{21,22,23} As well BPA strongly binds to transmembrane endoplasmic reticulum, G protein-coupled receptor 30 (GPR30) and estrogen-related receptor gamma (ERR γ).^{24,25} BPA can also activate peroxisome X receptor (PXR) and the aryl hydrocarbon receptor (AhR), often involved in cross-talk with steroid

receptors.^{26,27}

Many of these receptors play an important role in gene regulation, suggesting that BPA may influence normal differentiation and maturation processes especially during embryonic and fetal development.

The United States Environmental Protection Agency established a reference dose (RfD) for humans of 50 μg BPA/kg body weight (BW) day^{-1} , based on a thousand-fold reduction of the lowest observed adverse effect level (LOAEL) of 50 $\text{mg kg}^{-1}\text{BW day}^{-1}$.^{28,29} Studies indicate that daily human intake of BPA is less than 1 $\mu\text{g kg}^{-1}\text{BW day}^{-1}$, rendering the reference dose to be considered safe to humans.³⁰

However, other studies have shown that administration of low-dose BPA as 0.2 $\mu\text{g kg}^{-1}\text{BW day}^{-1}$ can reduce fertility and sperm production in male animals.^{31,32} At doses of 0.23–23 ng kg^{-1} BPA, the number of ERK-positive cerebellar cells increases and calcium ion signaling in pancreatic cells can be suppressed leading to diabetes mellitus.^{33,34,35}

To add, BPA is thought to elicit aneugenic effects by interfering with microtubule assembly, spindle apparatus function, chromosome segregation during mitosis^{36,37}, and disturbing DNA damage signaling pathways³⁸, thus affecting DNA stability, leading to potential carcinogenesis.

The content of Bisphenol A in amniotic fluid of pregnant women

Human pharmacokinetic data support rapid metabolism of free BPA to its BPA glucuronide (BPAG) and BPA sulfate (BPAS) metabolites, through UDP-glucuronyltransferase (*UGT2B15*) and sulfotransferase (*SULT1A1*) enzymes, resulting in faster urinary excretion of BPA in adults.^{39,40}

In comparison to adults, human fetuses and neonates have reduced capacity for chemical detoxification.^{41,42} Studies report that mammalian placenta presents with β -glucuronidase (*GUSB*) and steroid sulfatase (*STS*), breaking down inactive BPA metabolites to free BPA.^{43,44}

Performing liquid chromatography coupled with mass spectrometry (LC/MS) to compare levels of

conjugated and free BPA, in second and third trimester amniotic fluids, detected free BPA levels to comprise of 83% and 91% of total BPA, highlighting the role placental β -glucuronidase has on deconjugating BPA, thus exposing fetuses to even higher amount of free BPA and potentiating its adverse effects.⁴⁵

A considerate number of studies measured BPA in fetal cord blood^{46,47}, fetal liver^{48,49,50}, and amniotic fluids^{51,52}, on concentrations varying from 0.14 to 9.2, 1.3 to 50.5, and 0.36 to 5.62 ng/g, respectively.

Potential correlations between maternal and fetal blood in pregnant women, and between peripheral blood and peritoneal fluid in non-pregnant women, unveiled BPA levels ranging from non-detectable to 4.46ng/ml for maternal serum (MS) and from non-detectable to 4.60ng/ml in fetal serum (FS) of pregnant women. In the other hand, BPA levels in non-pregnant woman ranged from 1.30 to 8.17ng/ml in peripheral blood and from 0.19 to 13.45ng/ml in peritoneal fluid. Thus, positive correlation between maternal and fetal serum was found, highlighting a continuous distribution of BPA between the mother and fetus. Further, differing BPA concentration levels between pregnant and non-pregnant women reveal the role pregnancy has in underestimating the actual levels of BPA in blood.⁵³

A more complex study, using novel enzyme-linked immunosorbent assay (ELISA), compared BPA concentration on blood samples obtained from healthy premenopausal women, women with early and full-term pregnancy, ovarian follicular fluid, amniotic fluid and umbilical cord blood at full-term delivery.

Surprisingly, results revealed there was 5-fold higher concentration of BPA in amniotic fluid in comparison to other fluids, ranging between 8.3 +/- 8.7 ng/ml at 15-18 weeks of gestation. Findings suggest significant exposure during the prenatal period and accumulation of BPA in fetuses, which must be cautiously considered in evaluating the potential human exposure to endocrine-disrupting chemicals.⁵⁴

Differently, measuring BPA concentration levels in pregnant Korean women resulted in slightly higher BPA concentrations, ranging from non-detectable to 66.48 microg/L in mother serum, and from non-detectable to 8.86 microg/L in umbilical cords.⁵⁵ These higher BPA

rates in Korean pregnant women may be attributed to different geographical exposure rates to BPA, having in mind that BPA market in Asia raised 13% annually from 2000 to 2006.⁵⁶ So, different geographical exposure rates, should be considered when interpreting BPA concentration levels.

Moreover, a nested cross-sectional study revealed significant racial disparities in maternal and fetal BPA concentrations. African-Americans had 10-fold higher maternal serum BPA concentrations than Caucasians (30.13 vs 3.14ngml(-1); P=0.038), Hispanics had intermediate concentrations with a trend towards higher concentrations compared with Caucasians (24.46 vs 3.14ngml(-1); P=0.051) and Hispanics had higher fetal BPA concentrations than non-Hispanics (2.05 vs 0.35ngml(-1);P=0.025). These findings potentiate the immediate need to determine if such differences come from different levels of BPA exposure, fetal-placental transfer and its metabolism, or racial genetic variations.⁵⁷

It is crucial to know that measuring BPA as the end analyte might lead to inaccurate estimates, considering potential interferences from background sources during sample collection and analysis. Aglycone BPA and its primary conjugates as BPAG, BPAS, represent better candidates for biomarkers of BPA exposure, since they are not prone to external contamination and require in vivo metabolism.⁵⁸ To 2016 only ten studies reported analytical methods to measure BPA metabolites instead of just BPA. Research was limited by either lack of commercial or custom-synthesized BPA conjugates, or lack of labeled internal standards.^{59,60,61,62,63,64,65,66}

In order to achieve even more accurate and comprehensive representation of human exposure to endocrine disrupting chemicals, such as BPA, it is crucial for the future studies to consider cumulative effect of EDCs, knowing that humans are not exposed separately to just one EDC at a point of time.^{67,68,69}

BPA adverse health effects in fetuses and disease progression later in life

Exposure of rodent fetuses to Bisphenol A, at doses similar to environmental exposure, is found to cause postnatal estrogenic effects, as alteration of mammary gland morphology, detrimental long term effects in vagina and faster growth and puberty in females. As

well, reduced sperm production in males, increased prostate weight, and disruption of sexual differentiation in the brain was noticed.^{70,71,72,73,74,75,76}

Short time exposure to so considered “safe levels” of BPA, proved to have direct adverse effect on remodeling uterine spiral arteries, thus limiting blood supply to fetus⁷⁷, resulting to implantation failure, spontaneous abortion, recurrent miscarriages, or even leading to an increased risk of pre-eclampsia.^{78,79,80,81}

Studies support that women with detectable bisphenol A (BPA) concentrations have significantly higher risk of being infertile. La Rocca et al.⁸² found that the mean concentration of BPA was twice as high

in infertile than fertile women (10.6 vs. 4.8 ng ml⁻¹). Among infertile women, estrogen receptor alpha (ER α) and beta (ER β), androgen receptor (AR) and pregnane X receptor (PXR) were significantly expressed higher than in fertile patients, highlighting the distorting effect BPA has on these receptors.⁸³

Increased BPA concentrations are also reported to raise the occurrence of polycystic ovary syndrome (PCOS)⁸⁴, and are related to abnormalities in uterus morphology and endometriosis⁸⁵. Some other studies prove that BPA may cause atopic hyperplasia, uterine polyps or even cervical sarcoma and nipple adenoma.^{86,87,88}

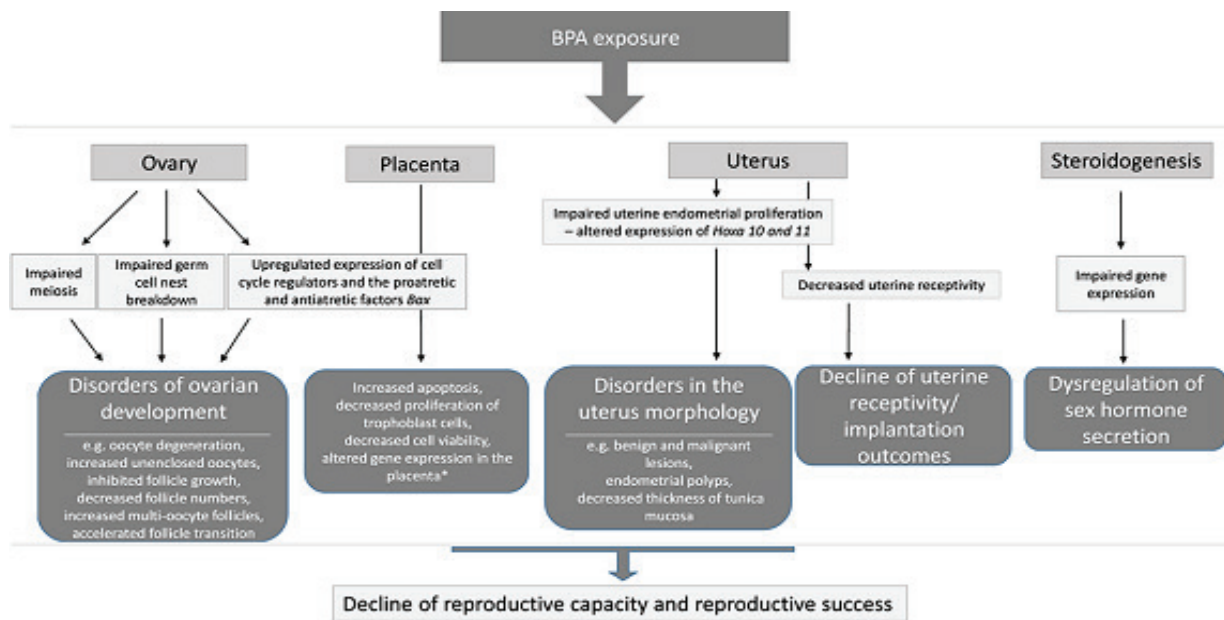


Figure 1: Effect of BPA on women reproductive processes. BPA, bisphenol A⁸⁹

In addition, BPA-treated testes contain mostly spermatogonia and spermatocytes with markedly less round spermatids, indicating signs of meiotic arrest. Neonatal BPA exposure disrupts meiosis during the first phase of spermatogenesis, due to inhibition

of *BOULE* (conserved key regulator for spermatogenesis expression and up-regulation of ER α/β expression in BPA-exposed developing testis)⁹⁰. This situation leads to increased presence of apoptotic cells in seminiferous tubules, sperm cells DNA damage and decreased sperm counts⁹¹.

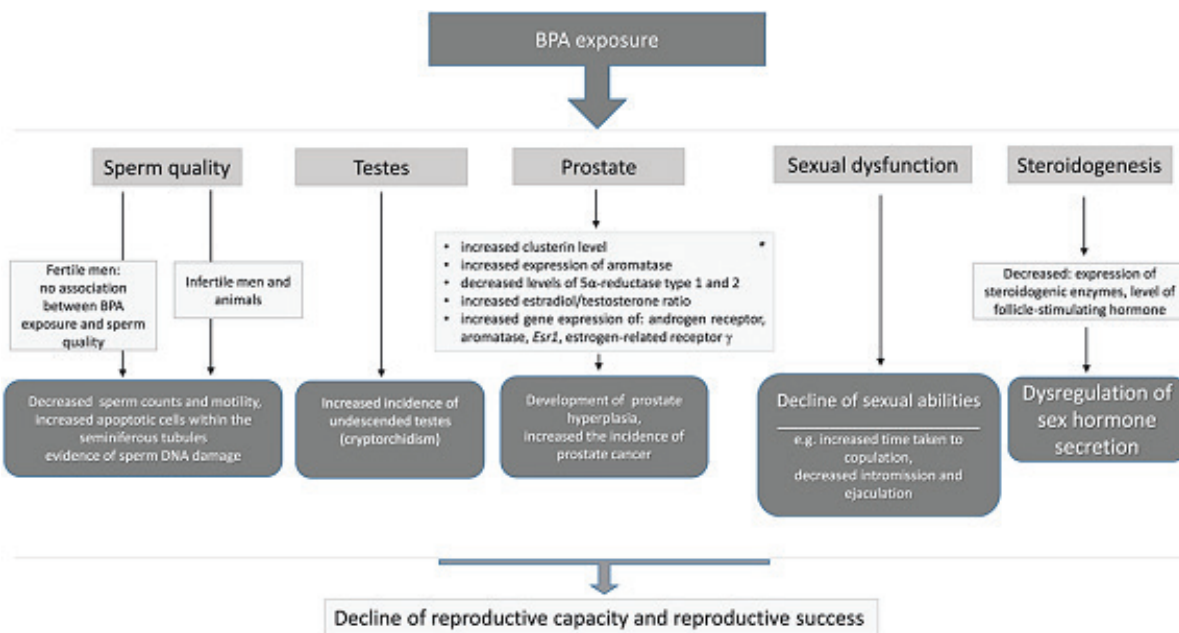


Figure 2: Effect of BPA on male reproductive processes. BPA, bisphenol A ⁸⁹

Rather than just having disruptive effects in reproductive tract, high BPA concentrations correlate with increased incidence of obesity and earlier puberty in females, altered physical and mental development in children, modified childhood behavior, cardiovascular disease and immune system dysfunction.^{92,93,94,95}

A ten year study, comparing BPA levels in amniotic fluid between pregnant mothers with normal and abnormal karyotype fetuses, highlighted that mothers with abnormal karyotype fetuses had higher levels of BPA concentrations in amniotic fluid, in comparison to pregnant women with normal karyotype fetuses. Therefore, these findings highlight the distorting potential BPA has in DNA stability and carcinogenesis induction.⁹⁶

Studies aiming to evaluate the role of BPA in carcinogenesis^{97,98} have indicated that exposure to BPA may increase the incidence of multiple cancers, as breast cancer^{99,100,101}, ovarian cancer^{102,103}, uterine cancer¹⁰⁴, prostate cancer^{105,106}, testicular cancer¹⁰⁷, and liver cancer.¹⁰⁸

Conclusions and Perspectives

Governmental restriction on BPA and general public concern regarding bisphenols adverse health effects, increased manufacturers interests on developing BPA

substitutes such as Bisphenol S (BPS) and Bisphenol F (BPF).¹⁰⁹ Due to their stability in sunlight and high temperature, they were initially thought to be safer alternatives.¹¹⁰ In contrast, in vitro studies found that BPS and BPF can elicit even greater estrogenic activities than BPA¹¹¹, can decrease cell viability, increase DNA damage and induce reproductive and neural toxicity.^{112,113,114,115,116}

In addition, having in mind BPA's accumulation potential, its distribution to fetal-placental unit, interspecies placental differences¹¹⁷, high BPA exposure in early weeks of gestations, racial disparities, varying geographical exposure rates, and BPA's cumulative effects with other EDCs, no endocrine disruptor dose should be assumed to be safe in pregnancy.^{118,119}

Therefore, increasing population's awareness around BPA adverse health effects through health education, and avoiding exposure to products containing BPA, remains the fastest and easiest way to limit BPA long life term adverse health outcomes.

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References

- Vandenberg L.N. et al. Human exposure to bisphenol A (BPA). *Reprod. Toxicol.* 2007. **24(2)**:139-77.
- Corrales J. et al. Global Assessment of Bisphenol A in the Environment. Review and Analysis of Its Occurrence and Bioaccumulation. *Dose Response.* 2015. **13(3)**
- Transparency Market Research Pvt. Ltd. Bisphenol A Market for Polycarbonates, Epoxy Resins and Other Applications – Global Industry Analysis, Size, Share, Growth and Forecast, 2013 – 2019. 2013. Retrieved from: <https://www.transparencymarketresearch.com/bisphenol-a-market.html>
- Rochester JR. Bisphenol A and human health: a review of the literature. *Reprod Toxicol.* 2013. **42**:132-55
- Geens T. et al. A review of dietary and non-dietary exposure to bisphenol-A. *Food Chem. Toxicol.* 2012. **50(10)**:3725-40
- U.S. Environmental Protection Agency. Bisphenol A alternatives in thermal paper. Final Report. 2014. Retrieved from: https://www.epa.gov/sites/production/files/2014-05/documents/bpa_final.pdf
- Staples CA. et al. A review of the environmental fate, effects, and exposures of bisphenol A. *Chemosphere.* 1998. **36**:2149–2173
- Heinonen J. et al. Bisphenol A accumulation in the freshwater clam *Pisidium amnicum* at low temperatures. *Arch Environ Contam Toxicol.* 2002. **43**:50–55
- Im J, Löffler FE. Fate of Bisphenol A in Terrestrial and Aquatic Environments. *Environ. Sci. Technol.* 2016. **50 (16)**:8403-16
- Valentino R. et al. Bisphenol A environmental exposure and the detrimental effects on human metabolic health: is it necessary to revise the risk assessment in vulnerable population. *Endocrinol. Invest.* 2016. **39**:259–263
- Howdeshell KL. et al. Bisphenol A is released from used polycarbonate animal cages into water at room temperature. *Environ Health Persp.* 2003. **111**:1180–1187
- Kang JH, Kito K, Kondo F. Factors influencing the migration of bisphenol A from cans. *J Food Prot.* 2003. **66**:1444–1447
- Olea N. et al. Estrogenicity of Resin-based Composites and Sealants Used in Dentistry. *Environ Health Persp.* 1996. **104**:298–305
- Geens T. et al. Assessment of human exposure to bisphenol A, triclosan and tetrabromobisphenol-A through indoor dust intake in Belgium. *Chemosphere.* 2009. **76**:755–760
- Soriano S. et al. Effects of Bisphenol A on ion channels: Experimental evidence and molecular mechanisms. *Steroids.* 2016. **111**:12-20
- Geens T, Neels H, Covaci A. Distribution of bisphenol-A, triclosan and n-nonylphenol in human adipose tissue, liver and brain. *Chemosphere.* 2012. **87(7)**:796-802.
- Ye X. et al. Measuring environmental phenols and chlorinated organic chemicals in breast milk using automated on-line column-switching-high performance liquid chromatography-isotope dilution tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2006. **2**:831(1-2):110-5
- Zhe Wang, Huiyu Liu, Sijin Liu. Low Dose Bisphenol A Exposure: A Seemingly Instigating Carcinogenic Effect on Breast Cancer. *Adv Sci (Weinh).* 2017. **4(2)**:1600248.
- Vandenberg L. N. et al. Bisphenol-A and the great divide: a review of controversies in the field of endocrine disruption. *Endocr Rev.* 2009. **30(1)**:75-95.
- Vandenberg L.N. et al. Human exposures to bisphenol A: mismatches between data and assumptions. *Rev Environ Health.* 2013. **28(1)**:37-58
- Gould JC, et al. Bisphenol A interacts with the estrogen receptor alpha in a distinct manner from estradiol. *Mol Cell Endocrinol.* 1998. **142(1-2)**:203–214.
- Kuiper GG. et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology.* 1998. **139(10)**:4252–4263.
- Moriyama K. et al. Thyroid hormone action is disrupted by bisphenol A as an antagonist. *J Clin Endocrinol Metab.* 2002. **87(11)**:5185–5190.
- Thomas P, Dong J. Binding and activation of the seven-transmembrane estrogen receptor GPR30 by environmental estrogens: a potential novel mechanism of endocrine disruption. *J Steroid Biochem Mol Biol.* 2006. **102(1-5)**:175–179.

25. Takayanagi S. et al. Endocrine disruptor bisphenol A strongly binds to human estrogen-related receptor gamma (ERRgamma) with high constitutive activity. *Toxicol Lett.* 2006. **167(2)**:95–105.
26. Sui Y. et al. Bisphenol A and its analogues activate human pregnane X receptor. *Environ Health Perspect.* 2012. **120(3)**:399–405.
27. Kruger T, Long M, Bonefeld-Jorgensen EC. Plastic components affect the activation of the aryl hydrocarbon and the androgen receptor. *Toxicology.* 2008. **246(2–3)**:112–123.
28. Bisphenol A. (CASRN 80-05-7). 1988. Retrieved from: http://www.epa.gov/nces/iris/iris_documents/documents/subst/0356 (accessed November 2016).
29. Welshons W. V. Large effects from small exposures. I. Mechanisms for endocrine-disrupting chemicals with estrogenic activity. *Environ Health Perspect.* 2003. **111(8)**:994–1006
30. Kang J. H., Kondo F., Katayama Y. Biodegradation or metabolism of bisphenol A: from microorganisms to mammals. *Toxicology.* 2006. **16;217(2-3)**:81-90
31. vom Saal F. S et al. A physiologically based approach to the study of bisphenol A and other estrogenic chemicals on the size of reproductive organs, daily sperm production, and behavior. *Toxicol Ind Health.* 1998. **14(1-2)**:239-60
32. Chitra K. C., Latchoumycandane C., Mathur P. P. Induction of oxidative stress by bisphenol A in the epididymal sperm of rats. *Toxicology.* 2003. **185(1-2)**:119-27
33. Zsarnovszky A. et al. Ontogeny of rapid estrogen-mediated extracellular signal-regulated kinase signaling in the rat cerebellar cortex: potent nongenomic agonist and endocrine disrupting activity of the xenoestrogen bisphenol A. *Endocrinology.* 2005. **146(12)**:5388-96
34. Alonso Magdalena P. Low doses of bisphenol A and diethylstilbestrol impair Ca²⁺ signals in pancreatic alpha-cells through a nonclassical membrane estrogen receptor within intact islets of Langerhans. *Environ Health Perspect.* 2005. **113(8)**:969-77
35. Provvizioso D. et al. Influence of Bisphenol A on Type 2 Diabetes Mellitus. *Int J Environ Res Public Health.* 2016. **13(10)**: 989
36. Ribeiro E, Ladeira C, Viegas S. EDCs Mixtures: A Stealthy Hazard for Human. *Health Toxic.* 2017. **5(1)**: 5.
37. Lehmann L, Metzler M. Bisphenol A and its methylated congeners inhibit growth and interfere with microtubules in human fibroblasts in vitro *Chem Biol Interact.* 2004. **147(3)**:273-85
38. Langie S. A. et al. Causes of genome instability: the effect of low dose chemical exposures in modern society. *Carcinogenesis.* 2015. **36 (1)**: 61-88
39. Hanioka N, Naito T, Narimatsu S. Human UDP-glucuronosyltransferase isoforms involved in bisphenol A glucuronidation. *Chemosphere.* 2008. **74(1)**:33–36.
40. Nishiyama T. et al. Sulfation of environmental estrogens by cytosolic human sulfotransferases. *Drug Metab Pharmacokinet.* 2002. **17(3)**:221–228.
41. Hines RN, McCarver DG. The ontogeny of human drug-metabolizing enzymes: phase I oxidative enzymes. *J Pharmacol. Exp. Ther.* 2002. **300(2)**:355–360.
42. McCarver DG, Hines RN. The ontogeny of human drug-metabolizing enzymes: phase II conjugation enzymes and regulatory mechanisms. *J Pharmacol Exp Ther.* 2002. **300(2)**:361–366.
43. Sperker B, Backman JT, Kroemer HK. The role of beta-glucuronidase in drug disposition and drug targeting in humans. *Clin Pharmacokinet.* 1997. **33(1)**:18–31.
44. Kriz L, Bicikova M, Hampl R. Roles of steroid sulfatase in brain and other tissues. *Physiol Res.* 2008. **57(5)**:657–668.
45. Edlow AG. et al. Fetal bisphenol A exposure: concentration of conjugated and unconjugated bisphenol A in amniotic fluid in the second and third trimesters. *Reprod Toxicol.* 2012. **34(1)**:1-7.
46. Schonfelder G. et al. Parent bisphenol A accumulation in the human maternal-fetal-placental unit. *Environ Health Persp.* 2002. **110**: A703–A707
47. Tan BLL, Mohd MA. Analysis of selected pesticides and alkylphenols in human cord blood by gas chromatograph-mass spectrometer. *Talanta.* 2003. **61**:385–391
48. Zhang J. et al. GC-MS analysis of bisphenol A in human placental and fetal liver samples. *J Chromatogr.* 2011. **B 879**:209–214
49. Cao XL. et al. Bisphenol A in human placental and fetal liver tissues collected from Greater Montreal area (Quebec) during 1998-2008. *Chemosphere.*

2012. **89**:505–511
50. Nahar MS. et al. Fetal Liver Bisphenol A Concentrations and Biotransformation Gene Expression Reveal Variable Exposure and Altered Capacity for Metabolism in Humans. *J Biochem Mol Toxic.* 2013.**27**:116–123
 51. Engel SM. et al. Xenobiotic phenols in early pregnancy amniotic fluid. *Reprod Toxicol.* 2006.**21**:110–112
 52. Chen F. et al. Distribution and accumulation of endocrine-disrupting chemicals and pharmaceuticals in wastewater irrigated soils in Hebei, China. *Environ Pollut.* 2011.**159**:1490–1498
 53. Aris A. Estimation of bisphenol A (BPA) concentrations in pregnant women, fetuses and nonpregnant women in Eastern Townships of Canada. *Reprod Toxicol.* 2014.**45**:8-13
 54. Ikezuki Y. et al. Determination of bisphenol A concentrations in human biological fluids reveals significant early prenatal exposure. *Hum Reprod.* 2002.**17(11)**:2839-41
 55. Lee YJ. et al. Maternal and fetal exposure to bisphenol A in Korea. *Reprod Toxicol.* 2008. **25(4)**:413-9
 56. Huang YQ et al. Bisphenol A (BPA) in China: a review of sources, environmental levels, and potential human health impacts. *Environ Int.* 2012.**42**:91-9
 57. Unal ER . et al. Racial disparity in maternal and fetal-cord bisphenol A concentrations. *J Perinatol.* 2012.**32(11)**:844-50.
 58. Andra S. et al. Recent advances in simultaneous analysis of bisphenol A and its conjugates in human matrices: exposure biomarker perspectives. *Sci Total Environ.* 2016. **572**: 770–781.
 59. Volkel W, Bittner N, Dekant W. Quantitation of bisphenol A and bisphenol A glucuronide in biological samples by high performance liquid chromatography-tandem mass spectrometry. *Drug Metab Dispos.* 2005.**33(11)**:1748–1757.
 60. Liao C, Kannan K. Determination of free and conjugated forms of bisphenol A in human urine and serum by liquid chromatography-tandem mass spectrometry. *Environmental Science and Technology.* 2012. **46(9)**:5003–5009.
 61. Nachman RM, et al. Serial Free Bisphenol A and Bisphenol A Glucuronide Concentrations in Neonates. *J Pediatr.* 2015.**167(1)**:64–69.
 62. Battal D, et al. Development and validation of an LC-MS/MS method for simultaneous quantitative analysis of free and conjugated bisphenol A in human urine. *Biomedical Chromatography.* 2014. **28(5)**:686–693.
 63. Provencher G, et al. Determination of bisphenol A, triclosan and their metabolites in human urine using isotope-dilution liquid chromatography-tandem mass spectrometry. *Journal of Chromatography A.* 2014.**1348**:97–104.
 64. Hauck ZZ, et al. Determination of bisphenol A-glucuronide in human urine using ultrahigh-pressure liquid chromatography/tandem mass spectrometry. *Rapid Commun Mass Spectrom.* 2016. **30(3)**:400–406.
 65. Lacroix MZ, et al. Simultaneous quantification of bisphenol A and its glucuronide metabolite (BPA-G) in plasma and urine: applicability to toxicokinetic investigations. *Talanta.* 2011. **85(4)**:2053–2059.
 66. Coughlin JL, Winnik B, Buckley B. Measurement of bisphenol A, bisphenol A -d-glucuronide, genistein, and genistein 4' - D-glucuronide via SPE and HPLC-MS/MS. *Analytical and Bioanalytical Chemistry.* 2011. **401(3)**:995–1002.
 67. Soto AM. et al. Developing a marker of exposure to xenoestrogen mixtures in human serum. *Environ Health Perspect.* 1997.**105(3)**:647–54.
 68. Vilahur N. et al. Prenatal exposure to mixtures of xenoestrogens and repetitive element DNA methylation changes in human placenta. *Environ Int.* 2014. **71**:81–7.
 69. Arrebola JP. et al. Predictors of the total effective xenoestrogen burden (TEXB) in human adipose tissue. A pilot study. *Reprod Toxicol.* 2012. **33(1)**:45–52.
 70. Howdeshell KL. et al. Exposure to bisphenol A advances puberty. *Nature.* 1999. **401(6755)**:763-4
 71. Kubo K. et al. Exposure to bisphenol A during the fetal and suckling periods disrupts sexual differentiation of the locus coeruleus and of behavior in the rat. *Neurosci Lett.* 2001. **304(1-2)**:73-6
 72. Markey CM. et al. In utero exposure to bisphenol A alters the development and tissue organization of the mouse mammary gland. *Biol Reprod.* 2001.**65(4)**:1215-23
 73. Nagel SC. et al. Relative binding affinity-serum modified access (RBA-SMA) assay

- predicts the relative in vivo bioactivity of the xenoestrogens bisphenol A and octylphenol. *Environ Health Perspect.* 1997. **105(1)**:70-6
74. Schönfelder G et al. Parent bisphenol A accumulation in the human maternal-fetal-placental unit. *Environ Health Perspect.* 2002. **110(11)**:A703-7
 75. Talsness Ch. et al. Components of plastic: experimental studies in animals and relevance for human health. *Philos Trans R Soc Lond B Biol Sci.* 2009. **364(1526)**: 2079–2096
 76. Welshons WV. et al. Low-dose bioactivity of xenoestrogens in animals: fetal exposure to low doses of methoxychlor and other xenoestrogens increases adult prostate size in mice. *Toxicol Ind Health.* 1999. **15(1-2)**:12-25
 77. Mülleret JE. et al. Bisphenol A exposure during early pregnancy impairs uterine spiral artery remodeling and provokes intrauterine growth restriction in mice. *Sci Rep.* 2018. **8**:9196.
 78. Sugiura-Ogasawara et al. Exposure to bisphenol A is associated with recurrent miscarriage. *Hum Reprod.* 2005. **(8)**:2325-9
 79. Shen Y et al. Higher urinary bisphenol A concentration is associated with unexplained recurrent miscarriage risk: evidence from a case-control study in eastern China. *PLoS One.* 2015. **26**:10(5)
 80. Ehrlich et al. Urinary bisphenol A concentrations and early reproductive health outcomes among women undergoing IVF. *Hum Reprod.* 2012. **27(12)**:3583-92
 81. Cantonwine et al. Urinary Concentrations of Bisphenol A and Phthalate Metabolites Measured during Pregnancy and Risk of Preeclampsia. *Environ Health Perspect.* 2016. **124(10)**:1651-1655
 82. La Rocca et al. Exposure to endocrine disrupters and nuclear receptor gene expression in infertile and fertile women from different Italian areas. *Int J Environ Res Public Health.* 2014. **11(10)**:10146-64
 83. Caserta D. et al. The influence of endocrine disruptors in a selected population of infertile women. *Gynecol Endocrinol.* 2013. **29(5)**:444-7
 84. Fernández M. et al. Neonatal exposure to bisphenol a and reproductive and endocrine alterations resembling the polycystic ovarian syndrome in adult rats. *Environ Health Perspect.* 2010. **118(9)**:1217-22
 85. Signorile PG. et al. Pre-natal exposure of mice to bisphenol A elicits an endometriosis-like phenotype in female offspring. *Gen Comp Endocrinol.* 2010. **168(3)**:318-25
 86. Adewale HB. et al. Neonatal bisphenol-a exposure alters rat reproductive development and ovarian morphology without impairing activation of gonadotropin-releasing hormone neurons. *Biol Reprod.* 2009. **81(4)**:690-9
 87. Fernandez SV, Russo J. Estrogen and xenoestrogens in breast cancer. *Toxicol Pathol.* 2010. **38(1)**:110-22
 88. Newbold RR, Jefferson WN, Padilla-Banks E. Prenatal exposure to bisphenol A at environmentally relevant doses adversely affects the murine female reproductive tract later in life. *Environ Health Perspect.* 2009. **117(6)**:879-85.
 89. Tomza Marciniak A. et al. Effect of bisphenol A on reproductive processes: A review of in vitro, in vivo and epidemiological studies. *Journal of Applied Toxicology.* 2017. **38 (1)**.
 90. Meina Xie et al. Neonatal bisphenol A exposure induces meiotic arrest and apoptosis of spermatogenic cells. *Oncotarget.* 2016. **1**:7(9)
 91. Meeker JD. et al. Semen quality and sperm DNA damage in relation to urinary bisphenol A among men from an infertility clinic. *Reprod Toxicol.* 2010. **30(4)**:532–539.
 92. Kaur K. et al. Bisphenol A induces oxidative stress and mitochondrial dysfunction in lymphoblasts from children with autism and unaffected siblings. *Free Radic Biol Med.* 2014. **76**:25-33
 93. Lang IA. et al. Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. *JAMA.* 2008. **300(11)**:1303–1310.
 94. Braun JM. et al. Impact of early-life bisphenol a exposure on behavior and executive function in children. *Pediatrics.* 2011. **128(5)**:873–882
 95. Robinson L., Miller R. The Impact of Bisphenol A and Phthalates on Allergy, Asthma, and Immune Function: A Review of Latest Findings . *Curr Environ Health Rep.* 2015. **2(4)**:379–387.
 96. Yamada H. et al. Maternal serum and amniotic fluid bisphenol A concentrations in the early second trimester. *Reprod Toxicol.* 2002. **16(6)**:735-9
 97. Keri R. et al. An evaluation of evidence for the

- carcinogenic activity of bisphenol A. *Reprod Toxicol.* 2007. **24(2)**:240-52
98. Gao H et al. Bisphenol A and hormone-associated cancers: current progress and perspectives. *Medicine (Baltimore)*.2015. **94(1)**:e211
 99. Ayyanan A. et al. Perinatal exposure to bisphenol a increases adult mammary gland progesterone response and cell number. *Mol Endocrinol.* 2011.**25(11)**:1915-23
 100. Murray T. J. et al. Induction of mammary gland ductal hyperplasias and carcinoma in situ following fetal bisphenol A exposure. *Reprod Toxicol.* 2007.**23(3)**:383-90
 101. Acevedo N. et al. Perinatally administered bisphenol a as a potential mammary gland carcinogen in rats. *Environ Health Perspect.* 2013.**121(9)**:1040-6
 102. Newbold RR, Jefferson WN, Padilla-Banks. Prenatal exposure to bisphenol a at environmentally relevant doses adversely affects the murine female reproductive tract later in life. *Environ Health Perspect.*2009. **117(6)**:879-85
 103. Fernandez M et al. Neonatal exposure to bisphenol A and reproductive and endocrine alterations resembling the polycystic ovarian syndrome in adult rats. *Environ Health Perspect.* 2010. **118(9)**:1217-22
 104. Newbold R. R., Jefferson W. N., Padilla-Banks E. Long-term adverse effects of neonatal exposure to bisphenol A on the murine female reproductive tract. *Reprod Toxicol.* 2007. **24(2)**:253-8
 105. Prins G. S. et al. Bisphenol A promotes human prostate stem-progenitor cell self-renewal and increases in vivo carcinogenesis in human prostate epithelium. *Endocrinology.* 2014. **155(3)**:805-17
 106. Prins G. S. et al. Serum bisphenol A pharmacokinetics and prostate neoplastic responses following oral and subcutaneous exposures in neonatal Sprague-Dawley rats. *Reprod Toxicol.* 2011.**31(1)**:1-9
 107. Nanjappa M. K., Simon L., Akingbemi B. T. The industrial chemical bisphenol A (BPA) interferes with proliferative activity and development of steroidogenic capacity in rat Leydig cells. *Biol Reprod.* 2012.**86(5)**:135
 108. Weinhouse C. et al. Dose-Dependent Incidence of Hepatic Tumors in Adult Mice following Perinatal Exposure to Bisphenol A. *Environ Health Perspect.* 2014. **122(5)**:485-491
 109. Rochester J. R., Bolden A. L. Bisphenol S and F: A Systematic Review and Comparison of the Hormonal Activity of Bisphenol A Substitutes. *Environ Health Perspect.* 2015.**123(7)**:643-50
 110. Qiu W. et al. Actions of Bisphenol A and Bisphenol S on the Reproductive Neuroendocrine System during Early Development in Zebrafish. *Endocrinology.* 2016. **157(2)**:636-47
 111. Chen D. et al. Bisphenol Analogues Other Than BPA: Environmental Occurrence, Human Exposure, and Toxicity-A Review. *Environ Sci Technol.* 2016.**50(11)**:5438-53
 112. Audebert M. et al. Use of the γ H2AX assay for assessing the genotoxicity of bisphenol A and bisphenol F in human cell lines. *Arch Toxicol.* 2011. **85(11)**:1463-73
 113. Michalowicz J., Mokra K., Bak A. Bisphenol A and its analogs induce morphological and biochemical alterations in human peripheral blood mononuclear cells (in vitro study). *Toxicol In Vitro.* 2015.**29(7)**:1464-72.
 114. Ji K. et al. Effects of bisphenol s exposure on endocrine functions and reproduction of zebrafish. *Environ Sci Technol.*2013. **47(15)**:8793-800.
 115. Švajger U, Dolenc MS, Jeras M. In vitro impact of bisphenols BPA, BPF, BPAF and 17 β -estradiol (E2) on human monocyte-derived dendritic cell generation, maturation and function. *Int Immunopharmacol.* 2016. 34:146-154
 116. Kinch C. D. et al. Low-dose exposure to bisphenol A and replacement bisphenol S induces precocious hypothalamic neurogenesis in embryonic zebrafish. *Proc Natl Acad Sci U S A.* 2015.112(5):1475-80
 117. Barry J, Anthony R. The Pregnant Sheep as a Model for Human Pregnancy. *Theriogenology.* 2008. **69(1)**:55-67.
 118. Shekhar S. et al. Detection of phenolic endocrine disrupting chemicals (EDCs) from maternal blood plasma and amniotic fluid in Indian population. *Gen Comp Endocrinol.* 2017. 241:100-107
 119. Vandenberg L. et al. Hormones and Endocrine-Disrupting Chemicals: Low-Dose Effects and Nonmonotonic Dose Responses. *Endocr Rev.* 2012. 33(3):378-455.